# **International Journal of Current Advanced Research**

ISSN: O: 2319-6475, ISSN: P: 2319-6505, Impact Factor: 6.614 Available Online at www.journalijcar.org Volume 7; Issue 10(C); October 2018; Page No. 15869-15877 DOI: http://dx.doi.org/10.24327/ijcar.2018.15877.2911



## HYDROXYAPATITE NANOPARTICLES AS DELIVERY SYSTEM IN COMBATING VARIOUS DISEASES

#### Ardhendu Kumar Mandal

Central Instrumentation Division CSIR-Indian Institute of Chemical Biology, India

ARTICLE INFO	ABSTRACT	

#### Article History:

Received 15<sup>th</sup> July, 2018 Received in revised form 7<sup>th</sup> August, 2018 Accepted 13<sup>th</sup> September, 2018 Published online 28<sup>th</sup> October, 2018

#### Key words:

Diseases; Hydroxyapatite nanoparticles; Mechanism of action; Immune responses; Delivery system Many people suffer from bone-related diseases such as osteoporosis, osteosarcoma, microbial infections and cancer throughout the world each year resulting in their deaths. Recently, hydroxyapatite nanoparticles have been emerged as a potent nanotechnology-based delivery system to treat several diseases. Owing to their nanosizes (<100 nm), high surface to volume ratio, easy of surface functionalization and suitable physico-chemical features such as bioactivity, biocompatibility, osteoinductivity and osteoconductivity, they are highly potent and favorable to generate cytotoxicity to kill the cells especially by the high uptake of calcium concentration within the cells. Moreover, their appropriate surface-modifications with ligands and other biomolecules may make them highly efficient carrier to deliver potent loaded-cargos to specific site of interest with a controllable and sustained manner leading to cellular destructions. The review demonstrates their synthesis, surface-functionalizations, mechanism of actions, immune responses and biomedical applications against various diseases to consider them as future nanotechnological delivery system.

Copyright©2018 Ardhendu Kumar Mandal. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## **INTRODUCTION**

Osteoporosis, a growing bony disease, is distinguished by reduced bone mass and micro-architectural bone tissue deterioration accompanied with increased bone fragility and subsequent susceptibility to fracture, and diagnosed when bone mineral density of a patient becomes 2.5 standard deviations or more below the average bone mass value for young healthy adults [Kanis and Kanis, 1994]. In adults, remodeling of bones is performed through a coordinated mechanism by which bone-resorbing osteoclasts remove old bones and boneforming osteoblasts mineralize and synthesize new bone matrix, while their disturbances in the physiological metabolic process cause reduced bone mass named as degenerative osteoporotic disorder [Beck et al., 2012; Riggs and Melton, 2015; Mackey and Whitaker, 2015]. Another neoplastic disorder is osteosarcoma characterized by malignant tumor of the skeleton [Abate et al., 2010; Pakos et al., 2009]. In this context, the growth of solid neoplasm is accompanied by neovascularization i.e. angiogenesis in tumor growth and progression and subsequent metastases supported by the overexpressions of vascular endothelial growth factor (VEGF) and matrix metalloproteinases [Carmeliet and Jain, 2000; Ferrare and Kerbel, 2005; Fukumura and Jain, 2007; Hao et al., 2015; Li et al., 2015; Ouyang et al., 2015; Shojaei, 2012; Holash et al., 1999; Vu et al., 1998].

\**Corresponding author:* Ardhendu Kumar Mandal Central Instrumentation Division CSIR-Indian Institute of Chemical Biology, India In another concern, mitochondrial dysfunction causes the generation of superoxide anion free radicals due to electron transport chain leakage resulting oxidative stress induction through reactive oxygen species (ROS) over-production [Turrens, 2003]. This ROS-production, inturn, causes several harmful consequences such as protein and lipid oxidations, mitochondrial RNA / DNA damages, Ca<sup>2+</sup>-dependent mitochondrial permeability transition pore activation and cytochrome c liberation following cellular apoptosis [Orrenius et al., 2007]. Both the mutations of mitochondrial proteins and oxidative stress trigger the cell death signaling cascade leading to organ damage, failure and disease development reflected on diabetes, cancer, neurodegenerative Alzheimer's and Parkinson's diseases, ischemia-reperfusion injury and heart failure [Taylor and Turnbull, 2005; Butterfield, 2002; Bayeva et al., 2013; Moreira et al., 2010; Weissig et al., 2007]. Furthermore, other criterion is the microbial infections associated with multidrug resistance and biofilm-development [Mandal, 2018]. These diseases generally develop when antioxidant defense system and innate and acquired immune system of the body become failure to overcome the inductive origin of disease development [Mandal, 2018].

In the past decades, global investigators have attempted to discover new solutions for improving treatments utilized for various bone related disorders, injuries, cancer and microbial infections. Their attention has been concentrated to the biomaterial field to create and develop new and improved ceramic biomaterials for tissue engineering where nanotechnology has restructured the conventional method of ceramic-usage in medical sciences. In general, patients suffer from drug resistance, insolubility, toxicity, bioavailability, enzymatic degradation, physiological barriers and specific target efficacy. In this concern, hydroxyapatite nanoparticles (HAp NPs,  $Ca(PO_4)_6(OH)_2$ ), the main component of the hard tissue such as bone and teeth, have attracted more attention in the field of biomedical applications owing to their exceptional characteristics such as favorable bioactivity, bioresorbablity, biocompatibility, biodegradability, osteoinductivity and osteoconductivity in treating bone related diseases, infective diseases, cancer and repair of hard tissue injury [Zhou and Lee, 2011; Danoux et al., 2014; Kobayashi and Murakoshi, 2014; Sooksaen et al., 2010; Zyman et al., 2013]. HAp NPs can exhibit significant cytotoxicity to diseased cells or organisms through mitochondria-dependent apoptotic induction emerging from oxidative stress and retardation of protein synthesis owing to their profuse nuclear and endoplasmic reticulum localizations as well as high intracellular concentrations of calcium ions (Ca<sup>2+</sup>) [Yuan et al., 2010; Meena et al., 2012; Qing et al., 2012; Chen et al., 2007; Xu et al., 2012; Han et al., 2014; Tang et al., 2014]. Moreover, HAp NPs may also be utilized as a carrier for protein, gene and drug delivery [Tada et al., 2010; Matsumoto et al., 2004; Uskokovic and Uskokovic, 2011]. These NPs can adsorb various chemicals onto their surfaces through electrostatic interactions, and owing to their soluble capability, the cargos may be liberated at specific targeted sites, while their solubility may be regulated by different substituted ions such as chloride, fluoride or carbonate, and low pH medium located in cancerous areas [Gomi et al., 1993]. Utilizing surfactants as the structure directing agent based on different synthesis techniques, pores were introduced into the diverse structure of HAp at the nano level to achieve desired loading and release profile of cargos [Al-Sokanee et al., 2009; Palazzo et al., 2005; Sadat-Shojaj et al., 2013]. In order to modulate HAp NPssurface, they may be coated with three layers of poly (allylamine), alginate and alendronate where outer alendronate layer is used as a targeting moiety to bind bone tissue and also as anti-resorptive drug to induce osteoconduction leading to increment of bone density in bone matrix for the treatment of osteolysis [Allen and Cullis, 2013]. HAp NPs loaded with appropriate active biomolecules may also be surfacefunctionalized with various ligands such as monoclonal antibody, sugar, protein, genes, lipids, poly ethylene glycol, poly (lactide-co-glycolide), poly (glycolide)-poly (ethylene glycol) and doped with rare-earth ions and metals to treat bone related diseases, cancer, metastases and infections [Dougall et al., 1999; Kim et al., 2016; Tokatlian and Segura, 2010; Giger et al., 2013; Hwang et al., 2016; Khajuria et al., 2016; Coelho et al., 2013; Han et al., 2008; Turner et al., 1989; Turner and Claringbold, 1991; Shi et al., 2009; Tai et al., 2013; Perry et al., 2012; Huang et al., 2008; Singh et al., 2015; Puljula et al., 2015].

This review demonstrates a survey of latest investigations regarding hydroxyapatite nanocomposites dealed with active ingredients-loaded preparatory methods, surface-modifications / encapsulations and doping with biomarkers accompanied with their targeting to specific diseased sites of interest crossing biological barriers for considering them as a highly promising delivery system against various diseases.

#### Synthesis of hydroxyapatite nanocomposites

HAp NPs are generally prepared by utilizing chemical precipitation, emulsion, hydrothermal and sol-gel methods. The most commonly used chemical precipitation methods are followed by any of the chemical reactions:

- 1.  $10 \text{ Ca}(\text{OH})_2 + 6 \text{ H}_3\text{PO}_4 \rightarrow \text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2 + 18 \text{ H}_2\text{O}$
- 2. 10 Ca(NO<sub>3</sub>)<sub>2</sub> + 6 (NH<sub>4</sub>)<sub>2</sub>HPO<sub>4</sub> + 2 H<sub>2</sub>O  $\rightarrow$  Ca<sub>10</sub>(PO<sub>4</sub>)<sub>6</sub>(OH)<sub>2</sub> + 12 NH<sub>4</sub>NO<sub>3</sub> + 8 HNO<sub>3</sub>

The basic common synthesis process involves the drop by drop addition of one phosphate reagent to the calcium reagent under continuous stirring, while the molar ratio of Ca/P maintained at HAp NPs is 1.67. In general, 5M aqueous  $Ca^{2+}$  ion solution is made by liquefying 0.03M Ca(NO<sub>3</sub>)<sub>2</sub>.4H<sub>2</sub>O in 6 mL distilled water. An aqueous 1500 µg / mL drug solution is separately prepared by dissolving drug into DMSO / Tween 20 and finally in distilled water. Afterthat, the solutions are stirred until a clear suspension is found. NH4OH is adjoined drop by drop to maintain the pH of Ca<sup>2+</sup> solution at 9 followed by the slow-addition of drug solution into this solution with continuous stirring. For maintaining the molar ratio of Ca to P at 1.67:1 in the reaction mixture, 0.018M (NH<sub>4</sub>)<sub>2</sub>HPO<sub>4</sub> is adjoined to Ca<sup>2+</sup>-drug solution following the re-adjustment of the pH of the reaction-mixture to 9 by drop by drop NH<sub>4</sub>OHaddition. The dissolution is kept for 24 h at room temperature and spun to void supernatant out. The precipitate is then cleansed with phosphate-buffered saline (PBS, pH 7.0) and deionized water three times for the removal of NO3 ions and loosely bound other components. 2M% Zn<sup>2+</sup> and Mg<sup>2+</sup>-doped HAp -drug NPs may be prepared by the adjoining of the needed amount of Zn(NO<sub>3</sub>)<sub>3</sub>.6H<sub>2</sub>O and Mg(NO<sub>3</sub>)<sub>3</sub>.6H<sub>2</sub>O respectively in the Ca<sup>2+</sup> aqueous solution. After cleansing, NPs are dried at room temperature. Porous HAp NPs may be prepared by various methods such as double emulsion [Shum et al., 2009], self-assembly [Saha et al., 2009; Xia et al., 2009; Ye et al., 2010; Huang et al., 2011], solvo-thermal [Ma and Zhu, 2009], hydrothermal [Wang et al., 2008; Ng et al., 2010; Guo et al., 2011; Guo et al., 2012], sol-gel [Dou et al., 2012] and spray drying [Sun et al., 2009]. During preparation of HAp NPs, an extensive regulated over size and morphology may be attained by modifications of experimental conditions [Zhang et al., 2009; Wuthier et al., 1985; Ren et al., 2013], the inclusion of surfactants [Cao et al., 2004] such as saccharides, cetyltrimethylammonium bromide (CTAB) and stearic acid, and chelating agents [Lopez et al., 1998; Ma, 2012] such as potassium sodium tartrate and trisodium citrate. Utilizing pore expander and cationic co-surfactant such as 1-dodecanethiol and CTAB respectively, large pore HAp NPs may also be synthesized [Bakhtiari et al., 2016].

Nanocrystalline hydroxyapatite doped with rare-earth samarium (Sm) element may be produced by adjusting the atomic ratio of Sm / [Sm+Ca] from 0% to 10% and [Ca+Sm] / P as 1.67. Samarium (III) nitrate hexahydrate and calcium nitrate tetrahydrate are liquefied in deionised water to get 300 mL Ca+Sm solution, while ammonium dihydrogen phosphate is liquefied in deionised water to form 300 mL P-containing solution. Then Ca+Sm solution is stirred at 100°C for 30 minutes, while pH of P-containing solution is set to 10 with NH<sub>3</sub> followed by stirring for 30 minutes. Then the P-containing solution is adjoined drop-wise into the Ca+Sm solution followed by stirring for 2 h, while the pH is adjusted

to 10. After that, the precipitate (Sm:HAp NPs) is cleansed several times with deionised water and dried at  $100^{\circ}C.$ 

Grafting or coating of HAp NPs with polymers suited with the copolymer / polymer matrix is also performed by several methods. For PGA-PEG coating of HAp NPs, 40 mg of this nanoparticles and 464 mg (4 mmol) of glycolide are dried overnight under 32 in Hg vacuum. Then HAp NPs and glycolide are dissoluted in 3 mL dimethylformamide (DMF), while glycolide, phosphazene base P2-t-Bu at 2M in THF is added to it. The reaction vessel is purged under N2 continuing 24 h reaction, while PGA coating-thickness is regulated by altering the ratio of glycolide and HAp NPs. Afterthat, 100 mg (0.02 mmol) of methoxy-poly (ethylene glycol)-isocyanate is adjoined directly into the reaction-mixture and kept on with for an additional 6 h under N<sub>2</sub> atmosphere. The yield is cleansed twice with DMF, and probable free polymer / monomer once with double distilled water through spinning at 5000 rpm for 5 min and lyophilized. The surface of HAp NPs may also be grafted through the reaction with their surface hydroxyls with poly (L-lactide-co-glycolide) (PLGA) to improve characteristics, dispersability and tensile strength [Song et al., 2013]. Furthermore, chitosan and carboxymethylated-chitosan may also be coated on the surface of HAp NPs to get higher solubility, viability and narrower size distribution [Muzzarelli et al., 2012; Berger et al., 2004; Kaya et al., 2015; Kaya et al., 2014; De Souza et al., 2009; Liang et al., 2004; Dumont et al., 2016: Barna et al., 2015]. In addition, to overcome microbial infection, biofilm and drug resistance, silver ions have been incorporated into HAp NPs-surface following co-precipitation method [Rameshbabu et al., 2007; Kim et al., 1998; Lim et al., 2015].

#### Characterizations of hydroxyapatite nanocomposites

The size and morphology of HAp nanocomposites may be monitored and determined by using transmission electron microscope, while their phase composition may be determined by X-ray diffraction analysis. The composition of nanocomposites may be analyzed by Fourier transform infrared (FT-IR) spectroscopy, while the zeta potential, hydrodynamic size and surface charge of particles in suspension may be measured by utilizing Zetasizer Nano ZS dynamic light scattering device.

#### Mechanism of action of hydroxyapatite nanoparticles

HAp NPs may be interiorized by cells via pinocytosis, phagocytosis, caveolae / clathrin -independent endocytosis and nonendocytosis pathways, while elevation of  $Ca^{2+}$  within cells and around endoplasmic reticulum hampers cellular Ca<sup>2+</sup> homeostasis causing oxidative stress mediated cytotoxicity leading to caspases- 9 and 3 -activated cellular apoptosis via the mitochondrial-dependent pathway, inhibition of protein synthesis by reducing the mRNA binding to its proper ribosomal binding site owing to their high adsorption capability for ribosome, and the cell cycle arrest in G0/G1 phase causing inhibition of cell proliferation, accompanied with necrosis and autophagic cell death [Zhao et al., 2011; Rothen et al., 2006; Monteith et al., 2007; Zhivotovsky and Orrenius, 2011; Meena et al., 2012; Xu et al., 2012; Han et al., 2014; Beland et al., 1979]. The overload of Ca<sup>2+</sup> by the exposure of HAp NPs is caused by their quick decadence in acidic cancer cells throughout intracellular translocation in phagolysosomal compartment containing hydrolases, while exorbitant extracellular Ca2+ influx, Ca2+ liberation from

intracellular storage or decreased  $Ca^{2+}$  efflux may also cause in raised  $Ca^{2+}$  level to produce cytotoxicity [Zhao *et al.*, 2011; Bloebaum *et al.*, 1998; Tang *et al.*, 2014; Orrenius *et al.*, 2003; Zhivotovsky and Orrenius, 2011].

## Hydroxyapatite nanoparticles as delivery vehicle

HAp NPs have been applied as a carrier to deliver varieties of therapeutics such as drugs, proteins, enzymes, antigens and genes [Thomas et al., 2015; Thomas et al., 2016]. HAp NPscoated liposomes have been utilized as efficacious drug delivery vectors for hydrophobic compounds such as indomethacin, a non-steroidal anti-inflammatory drug [Xu et al., 2007]. The osteoconductive HAp NPs loaded with bisphosphonate alendronate have been coated with layer-bylayer pH-responsive biopolymers poly (allylamine) and sodium alginate to treat osteoporosis for targeted drug delivery by providing superb biocompatibility and surface functionality via their COOH groups [Leu et al., 2006; Liang et al., 2012]. Based upon the surface functionalization of nanoparticles, especially by positively charged polymer chitosan coating, celecoxib has been encapsulated in the HAp NPs coated system for the treatment of colon cancer as well as tumor growth inhibition as an efficient vehicle for targeted drug delivery as positively charged HAp nanocomposites become highly accumulated into negatively charged cancer cells through electrostatic interactions [Yang and Hon, 2009; Li et al., 2009; Venkatesan et al., 2011]. As hydrophilic and negatively charged hyaluronic acid (HA) is a rising polymer to target the CD44 glycoprotein, expressed in tumor cells, some investigators have designed doxorubicin-loaded HAp NPs coated with HA to get efficient tumor targeting delivery [Zollen, 2011; Xiong et al., 2016]. Cancer may be dealt with cytotoxic free radicals and ROS through photodynamic therapy, but this technique has limitation owing to the low tissue piercing capability of visible light [Robertson et al., 2009; Klein et al., 2013]. In this context, researchers have reported metal-doped HAp NPs to utilize for cell apoptosis and *in vivo* tumor growth inhibition after irradiation with  $\gamma$ -rays as radiosensitizer metal ions give rise to significant ROS generation in cells for their damages [Kwatra et al., 2013; Matusiewicz, 2014; Chen et al., 2016]. Porous HAp NPs loaded with different antibiotics such as norflaxacin, ibuprofen, vancomycin exhibited high drug loading capacity and sustained liberation characteristics to the target site against infections [Melde and Stein, 2002; Tang et al., 2011; Ye et al., 2010]. Some researchers investigated in vitro desorption and adsorption of few anticancer drugs such as di (ethylenediamineplatinum) medronate, cisplatin and alendronate towards plate-shaped and needle-shaped porous HAp NPs, while the specific characteristics of the drugs and the morphology of the HAp NPs affected the desorption and adsorption kinetics of the drugs [Palazzo et al., 2007]. The other group of researcher prepared porous HAp NPs-surface functionalized and modified by using polyethylene glycol and folic acid with the attachment of anticancer paclitaxel drug, while the drug liberation profile exhibited an initial burst due to easily detachment from the NPs-surface, followed by a sustained release [Venkatasubbu et al., 2013].

# Biodistribution and elimination of hydroxyapatite nanoparticles

The biodistribution pattern of HAp NPs differs on their morphology, size, surface charge and surface chemistry along

with the modes of administration such as oral, intravenous, intraperitoneal, subcutaneous and intramuscular. HAp NPs may be decomposed to their consisting ions i.e.  $Ca^{2+}$ ,  $PO_4^{3-}$ and OH into the body after administration in an acidic environment due to the exchange of phosphate ions by hydrogen ions catalyzation from the NPs-surface into solution and recycled in bone construction [LeGeros, 1993; Christoffersen and Christoffersen, 1982]. Some particles may be endocytosed or phagocytosed by macrophages to accumulate into reticuloendothelial cells and become decomposed by lysosomal hydrolytic enzymes [Huang et al., 2004]. Smaller (<5 nm in size) HAp NPs are generally eliminated through urine, whereas larger NPs phagocytosed by macrophages undergo hepatobiliary excretion through feces [Mandal, 2018a]. Few researchers demonstrated that the intravenous administration of HAp NPs (40 nm and 200 nm in size) showed their major accumulation in liver and minor in spleen [Li and Huang, 2008; Ong et al., 2008]. Another group of investigators demonstrated that the HAp NPs (50-100 nm in size) conjugated to quantum dots showed their accumulation mainly in liver followed by spleen and other tissues elsewhere in the body [Guo et al., 2008]. Other group focused that rodlike HAp NPs (40-60 nm in size) loaded with plasmid DNA showed their accumulations in liver, kidney and brain tissue after intravenous administration [Zhu et al., 2004]. Few researchers also showed PEG-coated nanoparticles to accumulate in tumor cells through their leaky vascular endothelium due to enhanced permeation retention effect for longer blood circulation [Maeda, 2001].

#### Immune responses of hydroxyapatite nanoparticles

The administration of HAp NPs affects the rate of polymer matrix degradation causing inflammatory cell-mediated immune responses associated with antigen recognition by Tlymphocytes with consequent activations of the innate immune factors, mainly humoral macrophages and their derivatives, while the mature dendritic cells can activate the adaptive immune responses. The interactions with HAp NPs may cause death of macrophages implying their cytotoxic damage-effect by the suppression of the cellular immune responses. The immune responses occur primarily by the activation of inflammation and tissue damage accompanied by increased pro-inflammatory cytokines -secretions, secondly by the regulation of the secreted inflammatory cytokines governed by the macrophages, and thirdly by the induction of suppressive activity on the monocytes-differentiative energy to produce cytokines [Popova et al., 2011]. Some investigators studied in vitro cytokine levels while the exposure of HAp NPs with human monocytes THP-1 cells for 24 h exhibited increased levels of TNF- $\alpha$  and IL-1 $\beta$ , and increased IL-6 production in the co-cultures of THP-1cells and human umbilical vein endothelial cells (HUVECs) [Liu and Sun, 2014]. Few other researchers demonstrated the immunostimulatory potentials of HAp NPs in bone-marrow derived macrophages (BMDMs) and bone-marrow derived dendritic cells (BMDCs) and in vivo on their shape and size -dependent activations of the NLRP3 inflammosome and IL-1 $\beta$  secretion, while the smaller and needle -shaped HAp NPs significantly increased cytokinessecretion but not larger nanoparticles [Lebre et al., 2017].

## **CONCLUSIONS AND FUTURE PERSPECTIVES**

HAp NPs are reported to be a suitable candidate for biomedical usages due to their good biodegradability,

biocompatibility and bioactivity features, though their targeting and controlled contents-release to specific sites depends on the particles' synthesis, shape, size, compositions and surface modifications. Use of surfactants and ligands for the synthesis of HAp NPs to overcome aggregation or agglomeration, make them nano or meso -porous utilized for cargos- loading and release for specific targeting to cells owing to the NPs' high surface area and pore volume. Moreover, their doping with active compounds, rare-earth ions or metals makes them multifunctional not only for the treatments of low-bone density pathologies but also for the microbial, biofilm or cancer related diseases. The previous studies demonstrated that medium lethal dose (160 mg/kg) of HAp NPs to Wistar rats caused their deaths due to acute capillary blockage for the accumulation of NPs-aggregates [Aoki et al., 2000]. Therefore, the synthesis of the particles should be nano or meso -porous coated with cargos, ligands or doping with other materials to overcome the biological barriers, drug resistance, biofilm and toxic side effects and to deliver the specific active ingredients to the diseased site with sustained release manner. However, it is needed a thorough systematic study to synthesize HAp NPs with appropriate size, shape and functionalization with their biodistribution, pharmacokinetics, elimination, immune responses and efficacies to overcome systemic toxicity before going to clinics for consideration as delivery system for biomedical applications against different diseases.

#### Acknowledgement

This study was supported by the Council of Scientific and Industrial Research (CSIR), Government of India.

## References

- Abate, M.E., Longhi, A., Galletti, S., Ferrari, S. and Bacci, G. 2010. Non-metastatic osteosarcoma of the extremities in children aged 5 years or younger. Pediatric. Blood Cancer, 55:652-654.
- Allen, T.M. and Cullis, P.R. 2013. Liposomal drug delivery systems: From concept to clinical applications. Adv. Drug Deliv. Rev., 65:36-48.
- Al-Sokanee, Z., Toabi, A., Al-Assadi, M. and Alassadi, E.S. 2009. The drug release study of ceftriaxone from porous hydroxyapatite scaffolds. A.A.P.S. Pharm. Sci. Tech., 10:772-779.
- Aoki, H., Aoki, H., Kutsuno, T., Li, W. and Niwa, M. 2000. An *in vivo* study on the reaction of hydroxyapatite-sol injected into blood. *J. Mater. Sci. Mater. Med.*, 11:67-72.
- Bakhtiari, L., Javadpour, J., Rezaie, H.R., Erfan, M., Mazinani, B. and Aminian, A. 2016. Pore size control in the synthesis of hydroxyapatite nanoparticles: The effect of pore expander content and the synthesis temperature. Ceram. Int., 42:11259-11264.
- Barna, A.S., Ciobanu, G., Luca, C. and Luca, A.C. 2015. Nanohydroxyapatite-calcium fructoborate composites: Synthesis and characterization. Rev. Chim., 66:1618-1621.
- Bayeva, M., Gheorghiade, M. and Ardehali, H. 2013. Mitochondria as a therapeutic target in heart failure. J. Am. Coll. Cardiol., 61(6):599-610.
- Beck, G.R., Ha, S.W., Camalier, C.E., Yamaguchi, M., Li, Y., Lee, J.K. and Weitzmann, M.N. 2012. Bioactive silica-based nanoparticles stimulate bone-forming

osteoblasts, suppress bone-resorbing osteoclasts, and enhance bone mineral density *in vivo*. Nanomed. Nanotechnol. Biol. Med., 8:793-803.

- Beland, F.A., Dooley, K.L. and Casciano, D.A. 1979. Rapid isolation of carcinogen-bound DNA and RNA by hydroxyapatite chromatography. J. Chromatography A., 174:177-186.
- Berger, J., Reist, M., Mayer, J.M., Felt, O. and Gurny, R. 2004. Structure and interactions in chitosan hydrogels formed by complexation or aggregation for biomedical applications. Eur. J. Pharm. Biopharm., 57:35-52.
- Bloebaum, R.D., Lundeen, G.A., Bachus, K.N., Ison, I. and Hofmann, A.A. 1998. Dissolution of particulate hydroxyapatite in a macrophage organelle model. J. Biomed. Mater. Res., 40(1):104-114.
- Butterfield, D.A. 2002. Amyloid beta-peptide (1-42)induced oxidative stress and neurotoxicity: Implications for neurodegeneration in Alzheimer's disease brain. A review. Free Radic. Res., 36(12):1307-1313.
- Cao, M., Wang, Y., Guo, C., Qi, Y. and Hu, C. 2004. Preparation of ultrahigh-aspect-ratio hydroxyapatite nanofibers in reverse micelles under hydrothermal conditions. Langmuir., 20:4784-4786.
- Carmeliet, P. and Jain, R.K. 2000. Angiogenesis in cancer and other diseases. Nature, 407:249-257.
- Chen, M.H., Hanagata, N., Ikoma, T., Huang, J.Y., Li, K.Y., Lin, C.P. and Lin, F.H. 2016. Hafnium-doped hydroxyapatite nanoparticles with ionizing radiation for lung cancer treatment. Acta. Biomater., 37:165-173.
- Chen, X.J., Deng, C.S., Tang, S.L. and Zhang, M. 2007. Mitochondria dependent apoptosis induced by nanoscale hydroxyapatite in human gastric cancer SGC-7901 cells. Biol. Pharm. Bull., 30(1):128-132.
- Christoffersen, J. and Christoffersen, M.R. 1982. Kinetics of dissolution of calcium hydroxyapatite: V. The acidity constant for the hydrogen phosphate surface complex. J. Cryst. Growth, 57:21-26.
- Coelho, J., Hussain, N.S., Gomes, P.S., Garcia, M.P., Lopes, M.A., Fernandes, M.H. and Santos, J.D. 2013.
  Development and characterization of lanthanides doped hydroxyapatite composites for bone tissue application. In current Trends on Glass and Ceramic Materials. Scopus, pp.87-115.
- Danoux, C.B., Barbieri, D., Yuan, H., de Bruijin, J.D., van Blitterswijra, C.A. and Habibovic, P. 2014. *In vitro* and *in vivo* bioactivity assessment of a polylactic acid / hydroxyapatite composite for bone regeneration. Biomat., 4:e27664.
- DeSouza, C.J.E., Pereira, M.M. and Mansur, H.S. 2009. Properties and biocompatibility of chitosan films modified by blending with PVA and chemically crosslinked. *J. Mater. Sci. Mater. Med.*, 20:553-561.
- Dou, Y., Cai, S., Ye, X., Xu, G., Hu, H. and Ye, X. 2012. Preparation of mesoporous hydroxyapatite films used as biomaterials via sol-gel technology. J. Sol-Gel Sci. Technol., 61:126-132.
- Dougall, W.C., Glaccum, M., Charrier, K., Rohrbach, K., Brasel, K., Smedt, TD., Daro, E., Smith, J., Tometsko, M.E., Maliszewski, C.R., Armstrong, A., Shen, V., Bain, S., Cosman, D., Anderson, D., Morrissey, P.J., Peschon, J.J. and Schuh, J. 1999. RANK is essential for osteoclast and lymph node development. Gene Dev., 13(18):2412-2424.

- Dumont, V.C., Mansur, A.A.P., Carvalho, S.M., Medeiros, B.F.G.L., Pereira, M.M. and Mansur, H.S. 2016. Chitosan and carboxymethyl-chitosan capping ligands: Effects on the nucleation and growth of hydroxyapatite nanoparticles for producing biocomposite membranes. Mater. Sci. Eng. C., 59:265-277.
- Ferrare, N. and Kerbel, R.S. 2005. Angiogenesis as a therapeutic target. Nature, 438:967-974.
- Fukumura, D. and Jain, R.K. 2007. Tumor microvasculature and microenvironment: Targets for anti-angiogenesis and normalization. Microvasc. Res., 74:72-84.
- Giger, E.V., Castagner, B. and Leroux, J.C. 2013. Biomedical applications of bisphosphonates. J. Control. Release, 167; 175-188.
- Gomi, K., Lowenberg, B., Shapiro, G. and Davies, J.E. 1993. Resorption of sintered synthetic hydroxyapatite by osteoclasts in vitro. Biomat., 14(2):91-96.
- Guo, Y., Shi, D., Lian, J., Dong, Z., Wang, W., Cho, H., Liu, G., Wang, L. and Ewing, R. 2008. Quantum dot conjugated hydroxyapatite nanoparticles for *in vivo* imaging. Nanotechnol., 19:175102-175107.
- Guo, Y.P., Yao, Y., Ning, C.Q., Guo, Y.J. and Chu, L.F. 2011. Fabrication of mesoporous carbonated hydroxyapatite microspheres by hydrothermal method. Mater. Lett., 65:2205-2208.
- Guo, Y.P., Yao, Y.B., Guo, Y.J. and Ning, C.Q. 2012. Hydrothermal fabrication of mesoporous carbonaqted hydroxyapatite microspheres for a drug delivery system. Microporous Mesoporous Mater., 155:245-251.
- Han, Y., Li, S., Cao, X., Yuan, L., Wang, Y., Yin, Y., Qiu, T., Dai, H. and Wang, X. 2014. Different inhibitory effect and mechanism of hydroxyapatite nanoparticles on normal cells and cancer cells *in vitro* and *in vivo*. Sci. Rep., 4:7134.
- Han, Y.J., Loo, S.C.J., Phung, N.Y., Boey, F. and Ma, J. 2008. Controlled size and morphology of EDTMPdoped hydroxyapatite nanoparticles as model for <sup>153</sup>Samarium-EDTMP doping. J. Mater. Sci. Mater. Med., 19(9):2993-3003.
- Hao, S., Yan, Y., Ren, X., Xu, Y., Chen, L. and Zhang, H. 2015. Candesartan-graft-polyethyleneimine cationic micelles for effective co-delivery of drug and gene in anti-angiogenic lung cancer therapy. Biotechnol. Bioproc. E., 20:550-560.
- Holash, J., Wiegand, S. and Yancopoulos, G. 1999. New model of tumor angiogenesis: Dynamic balance between vessel regression and growth mediated by angiopoietins and VEGF. Oncogene, 18(38):5356-5362.
- Huang, J., Best, S.M., Bonfield, W., Brooks, R.A., Rushton, N., Jayasinghe, S.N. and Edirisingho, M.J. 2004. In vitro assessment of the biological response to nanosized hydroxyapatite. J. Mater. Sci. Mater. Med., 15:441-445.
- Huang, Y.T., Imura, M., Nemoto, Y., Cheng, C.H. and Yamauchi, Y. 2011. Block-copolymer-assisted synthesis of hydroxyapatite nanoparticles with high surface area and uniform size. Sci. Technol. Adv. Mater., 12:045005.
- Huang, Y.X., Ren, J., Chen, C., Ren, T.B. and Zhou, X.Y. 2008. Preparation and properties of poly (lactide-coglycolide) (PLGA) / nano-hydroxyapatite (NHA) scaffold by thermally induced phase separation and

rabbit MSCs culture on scaffolds. J. Biomater. Appl., 22:409-432.

- Hwang, S.J., Lee, J.S., Ryu, T.K., Kang, R.H., Jeong, K.Y., Jun, D.R., Koh, J.M., Kim, S.E. and Choi, S.W. 2016. Alendronate-modified hydroxyapatite nanoparticles for bone-specific dual delivery of drug and bone mineral. Macromol. Res., 24(7):623-628.
- Kanis, J.A. and Kanis, J.A. 1994. Assessment of fracture risk and its application to screening for postmenopausal osteoporosis: Synopsis of a WHO report. Osteopor. Int., 4:368-381.
- Kaya, M., Baran, T., Asan, O.M., Cakmak, Y.S., Tozak, K.P., Mol, A., Mentes, A. and Sezen, G. 2015. Extraction and characterization of chitin and chitosan with antimicrobial and antioxidant activities from cosmopolitan Orthoptera species (Insecta). Biotechnol. Bioproc. Eng., 20:168-179.
- Kaya, M., Cakmak, Y.S., Baran, T., Asan, O.M., Mentes, A. and Tozak, K.O. 2014. New chitin, chitosan, and Ocarboxymethyl chitosan sources from resting eggs of *Daphnia longispina* (Crustacea); with physicochemical characterization, and antimicrobial and antioxidant activities. Biotechnol. Bioproc. E., 19:58-69.
- Khajuria, D.K., Disha, C., Vasireddi, R., Razdan, R. and Mahapatra, D.R. 2016. Risedronate / zinchydroxyapatite based nanomedicine for osteoporosis. Mat. Sci. Eng. C. Mater. Biol. Appl., 63:78-87.
- Kim, T., Singh, R.K., Kang, M.S., Kim, J.H. and Kim, H.W. 2016. Inhibition of osteoclastogenesis through siRNA delivery with tunable mesoporous bioactive nanocarriers. Acta. Biomat., 29:352-364.
- Kim, T.N., Feng, Q.L., Kim, J.O., Wu, J., Wang, H., Chen, G.C. and Cui, F.Z. 1998. Antimicrobial effects of metal ions (Ag<sup>+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>) in hydroxyapatite. *J. Mater. Sci. Mater. Med.*, 9:129-134.
- Klein, S., Dell, A.M.L., Wegmann, M., Distel, L.V., Neuhuber, W., Gonzalez, M.C. and Kryschi, C. 2013. Oxidized silicon nanoparticles for radiosensitization of cancer and tissue cells. Biochem. Biophys. Res. Commun., 434:217-222.
- Kobayashi, S. and Murakoshi, T. 2014. Characterization of mechanical properties and bioactivity of hydroxyapatite
   / β-tricancium phosphate composites. Adv. Compos. Mater., 23:163-177.
- Kwatra, D., Venugopal, A. and Anant, S. 2013. Nanoparticles in radiation therapy: A summary of various approaches to enhance radiosensitization in cancer. Transl. cancer Res., 2:330-342.
- Lebre, F., Sridharan, R., Sawkins, M.J., Kelly, D.J., O'Brien, F.J. and Lavelle, E.C. 2017. The shape and size of hydroxyapatite particles dictate inflammatory responses following implantation. Sci. Rep., 7:2922.
- LeGeros, R.Z. 1993. Biodegradation and bioresorption of calcium phosphate ceramics. Clin. Mater., 14:65-88.
- Leu, C.T., Luegmayr, E., Freedman, L.P., Rodan, G.A. and Reszka, A.A. 2006. Relative binding affinities of bisphosphonates for human bone and relationship to antiresponsive efficacy. Bone, 38:628-636.
- Li, F., Li, J., Wen, X., Zhou, S., Tong, X., Su, P., Li, H. and Shi, D. 2009. Anti-tumor activity of paclitaxel-loaded chitosan nanoparticles: An *in vitro* study. Mater. Sci. Eng. C., 29:2392-2397.

- Li, M., Li, Y., Huang, X. and Lu, X. 2015. Captoprilpolyethyleneimine conjugate modified gold nanoparticles for co-delivery of drug and gene in antiangiogenesis breast cancer therapy. J. Biomater. Sci. Polym. Ed., 26(13):813-827.
- Li, S. and Huang, L. 2008. Pharmacokinetics and biodistribution of nanoparticles. Mol. Pharm., 5:496-504.
- Liang, P., Zhao, Y., Shen, Q., Wang, D.J. and Xu, D.F. 2004. The effect of carboxymethyl-chitosan on the precipitation of calcium carbonate. *J. Cryst. Growth*, 261:571-576.
- Liang, Y.H., Liu, C.H., Liao, S.H., Lin, Y.Y., Tang, H.W., Liu, S.Y., Lai, I.R. and Wu, K.C. 2012. Cosynthesis of cargo-loaded hydroxyapatite / alginate core-shell nanoparticles (HAP@Alg) as pH-responsive nanovehicles by a pre-gel method. A.C.S. Appl. Mater. Interfaces, 4:6720-6727.
- Lim, P.N., Chang, L. and Thian, E.S. 2015. Development of nanosized silver-substituted apatite for biomedical applications: A review. Nanomed. N.B.M., 11:1331-1344.
- Liu, X. and Sun, J. 2014. Potential pro-inflammatory effects of hydroxyapatite nanoparticles on endothelial cells in a monocyte-endothelial cell co-culture model. Int. J. Nanomed., 9:1261-1273.
- Lopez, M.A., Gomez, M.J., Rodriguez, C.R. 1998. Nanosized hydroxyapatite precipitation from homogenous calcium / citrate / phosphate solutions using microwave and conventional heating. Adv. Mater., 10:49-53.
- Ma, M.G. 2012. Hierarchically nanostructured hydroxyapatite: Hydrothermal synthesis, morphology control, growth mechanism, and biological activity. Int. J. Nanomed., 7:1781-1791.
- Ma, M.G. and Zhu, J.F. 2009. Solvothermal synthesis and characterizationof hierarchically nanostructured hydroxyapatite hollow spheres. Eur. J. Inorg. Chem., 36:5522-5526.
- Mackey, P.A. and Whitaker, M.D. 2015. Osteoporosis: A therapeutic update. J. Nurse. Pract., 11:1011-1017.
- Maeda, H. 2001. The enhanced permeability and retention (EPR) effect in tumor vasculature: The key role of tumor-selective macromolecular drug targeting. Adv. Enzyme Regul., 41:189-207.
- Mandal, A.K. 2018. Zinc oxide nanoparticles as delivery system to combat diseases. Int. J. Current Adv. Res., 7(5D):12469-12478.
- Mandal, A.K. 2018a. Mesoporous silica nanoparticles as delivery system against diseases. Int. J. Recent Sci. Res., 9(9):28817-28825.
- Matsumoto, T., Okazaki, M., Inoue, M., Yamaguchi, S., Kusunose, T., Toyonaga, T., Hamada, Y. and Takahashi, J. 2004. Hydroxyapatite particles as a controlled release carrier of protein. Biomat., 25(17):3807-3812.
- Matusiewicz, H. 2014. Potential release of *in vivo* trace metals from metallic medical implants in the human body: From ions to nanoparticles A systematic analytical review. Acta. Biomater., 10:2379-2403.
- Meena, R., Kesari, K.K., Rani, M. and Paulraj, R. 2012. Effects of hydroxyapatite nanoparticles on proliferation

and apoptosis of human breast cancer cells (MCF-7). J. Nanopart. Res., 14(2):712.

- Melde, B.J. and Stein, A. 2002. Periodic macroporous hydroxyapatite-containing calcium phosphates. Chem. Mater., 14(8):3326-3331.
- Monteith, G.R., McAndrew, D., Faddy, H.M. and Roberts, T.S.J. 2007. Calcium and cancer: Targeting Ca<sup>2+</sup> transport. Nat. Rev. Cancer, 7(7):519-530.
- Moreira, P.I., Zhu, X., Wang, X., Lee, H.G., Nunomura, A., Petersen, R.B., Perry, G. and Smith, M.A. 2010. Mitochondria: A therapeutic target in neurodegeneration. Biochim. Biophys. Acta., 1802(1):212-220.
- Muzzarelli, R.A.A., Boudrant, J., Meyer, D., Manno, N., Demarchis, M. and Paoletti, M.G. 2012. Current views on fungal chitin / chitosan, human chitinases, food preservation, glucans, pectins and inulin: A tribute to Henri Braconnot, precursor of the carbohydrate polymers science, on the chitin bicentennial. Carbohydr. Polym., 87: 995-1012.
- Ng, S.X., Guo, J., Ma, J. and Loo, S.C.J. 2010. Synthesis of high surface area mesostructured calcium phosphate particles. Acta. Biomater., 6:3772-3781.
- Ong, H., Loo, J., Boey, F., Russel, S., Ma, J., Peng, K. 2008. Exploiting the high-affinity phosphonate-hydroxyapatite nanoparticles interaction for delivery of radiation and drugs. J. Nanopart. Res., 10:141-150.
- Orrenius, S., Gogvadze, V. and Zhivotovsky, B. 2007. Mitochondrial oxidative stress: Implications for cell death. Annu. Rev. Pharmacol. Toxicol., 47:143-183.
- Orrenius, S., Zhivotovsky, B. and Nicotera, P. 2003. Regulation of cell death: The calcium-apoptosis link. Nat. Rev. Mol. Cell. Biol., 4(7):552-565.
- Ouyang, Q., Duan, Z., Jiao, G. and Lei, J. 2015. A biomimic reconstituted high-density-lipoprotein-based drug and p53 gene co-delivery system for effective antiangiogenesis therapy of bladder cancer. Nanoscale Res. Lett., 10:965.
- Pakos, E.E., Nearchou, A.D. and Grimer, R.J. 2009. Prognostic factors and outcomes for osteosarcoma: An International Collaboration. Eur. J. Cancer, 45:2367-2375.
- Palazzo, B., Iafisco, M., Laforgia, M., Margiotta, N., Natile, G., Bianchi, C.L., Walsh, D., Mann, S. and Roveri, N. 2007. Biomimetic hydroxyapatite-drug nanocrystals as potential bone substitutes with antitumor drug delivery properties. Adv. Funct. Mater., 17:2180-2188.
- Palazzo, B., Sidoti, M.C., Roveri, N., Tampieri, A., Sandri, M., Bertolazzi, L., Galbusera, F., Dubini, G., Vena, P. and Contro, R. 2005. Controlled drug delivery from porous hydroxyapatite grafts: An experimental and theoretical approach. Mater. Sci. Eng. C., 25:207-213.
- Perry, J.L., Reuter, K.G., Kai, M.P., Herlihy, K.P., Jones, S.W., Luft, J.C., Napier, M., Bear, J.E. and DeSimone, J.M. 2012. PEGylated PRINT nanoparticles: The impact of PEG density on protein binding, macrophage association, biodistribution, and pharmacokinetics. Nano Lett., 12(10):5304-5310.
- Popova, A., Kzhyshkowska, J., Nurgazieva, D., Goerdt, S. and Gratchev, A. 2011. Pro- and anti- inflammatory control of M-CSF mediated macrophage differentiation. Immunobiol., 216:164-172.

- Puljula, E., Yurhanen, P., Vepsalainen, J., Monteil, M., Lecouvey, M. and Weisell, J. 2015. Structural requirements for bisphosphonate binding on hydroxyapatite: NMR study of bisphosphonate partial esters. A.C.S. Med. Chem. Lett., 6:397-401.
- Qing, F.Z., Wang, Z., Hong, Y.L., Liu, M., Guo, B., Luo, H.R. and Zhang, X.D. 2012. Selective effects of hydroxyapatite nanoparticles on osteosarcoma cells and osteoblasts. J. Mater. Sci. Mater. Med., 23(9):2245-2251.
- Rameshbabu, N., Sampath, K.T.S., Prabhakar, T.G., Sastry, V.S., Murty, K.V.G.K. and Prasad, R.K. 2007. Antibacterial nanosized silver substituted hydroxyapatite: Synthesis and characterization. J. Biomed. Mater. Res. A., 80:581-591.
- Ren, F., Leng, Y., Ding, Y. and Wang, K. 2013. Hydrothermal growth of biomimetric carbonated apatite nanoparticles with tunable size, morphology and ultrastructure. Cryst. Eng. Commun., 15:2137-2146.
- Riggs, B.L. and Melton, L.J. 2015. The worldwide problem of osteoporosis: Insights afforded by epidemiology. Bone, 17:505S-511S.
- Robertson, C.A., Evans, D.H. and Abrahamse, H. 2009. Photodynamic therapy (PDT): A short review on cellular mechanisms and cancer research applications for PDT. J. Photochem. Photobiol. B., 96:1-8.
- Rothen, R.B.M., Schuerch, S., Haenni, B., Kapp, N. and Gehr, P. 2006. Interaction of fine particles and nanoparticles with red blood cells visualized with advanced microscopic techniques. Environ. Sci. Technol., 40(14): 4353-4359.
- Sadat-Shojaj, M., Khorasani, M.T., Dinpanah-Khoshdargi, E., Jamshidi, A. 2013. Synthesis methods for nanosized hydroxyapatite with diverse structures. Acta. Biomater., 9(8):7591-7621.
- Saha, S.K., Banerjee, A., Banerjee, S. and Bose, S. 2009. Synthesis of nanocrystalline hydroxyapatite using surfactant template systems: Role of templates in controlling morphology. Mater. Sci. Eng. C., 29:2294-2301.
- Shi, X., Wang, Y., Ren, L., Gong, Y. and Wang, D.A. 2009. Enhancing alendronate release from a novel PLGA / hydroxyapatite microspheric system for bone repairing applications. Pharm. Res., 26:422-430.
- Shojaei, F. 2012. Anti-angiogenesis therapy in cancer: Current challenges and future perspectives. Cancer Lett., 320:130-137.
- Shum, H.C., Bandyopadhyay, A., Bose, S. and Weitz, D.A. 2009. Double emulsion droplets as microreactors for synthesis of mesoporous hydroxyapatite. Chem. Mater., 21:5548-5555.
- Singh, T., Kaur, V., Kumar, M., Kaur, P., Murthy, R.S. and Rawal, R.K. 2015. The critical role of bisphosphonates to target bone cancer metastasis: An overview. J. Drug Target., 23:1-15.
- Song, X.F., Ling, F.G. and Chen, X.S. 2013. Grafting polymerization of L-lactide on the surface of hydroxyapatite nanoparticles. Acta. Polym. Sin., 1:95-101.
- Sooksaen, P., Jumpanoi, N., Suttiphan, P. and Kimchaiyong, E. 2010. Crystallization of nano-sized hydroxyapatite via wet chemical process under strong alkaline conditions. Sci. J. U.B.U., 1:20-27.

- Sun, R., Chen, K. and Lu, Y. 2009. Fabrication and dissolution behavior of hollow hydroxyapatite microspheres intended for controlled drug release. Mater. Res. Bull., 44:1939-1942.
- Tada, .S, Chowdhury, E.H., Cho, C.S. and Akaike, T. 2010. pH-sensitive carbonate apatite as an intracellular protein transporter. Biomat., 31(6):1453-1459.
- Tai, I.C., Fu, Y.C., Wang, C.K., Chang, J.K. and Ho, M.L. 2013. Local delivery of controlled release simvastatin / PLGA / HAp microspheres enhances bone repair. Int. J. Nanomed., 8:3895-3904.
- Tang, Q.L., Zhu, Y.J., Wu, J., Chen, F. and Cao, S.W. 2011. Calcium phosphate drug nanocarriers with ultrahigh and adjustable drug-loading capacity: One-step synthesis, *in situ* drug loading and prolonged drug release. Nanomed., 7:428-434.
- Tang, W., Yuan, Y., Liu, C.S., Wu, Y.Q., Lu, X. and Qian, J.C. 2014. Differential cytotoxicity and particle action of hydroxyapatite nanoparticles in human cancer cells. Nanomed., 9(3):397-412.
- Taylor, R.W. and Turnbull, D.M. 2005. Mitochondrial DNA mutations in human disease. Nat. Rev. Genet., 6(5):389-402.
- Thomas, S.C., Harshita, Mishra, P.K. and Talegaonkar, S. 2015. Ceramic nanoparticles: Fabrication methods and applications in drug delivery. Curr. Pharm. Des., 21:6165-6188.
- Thomas, S.C., Sharma, H., Rawat, P., Verma, A., Leekha, A., Kumar, V., Tyagi, A., Gurjar, B.S., Iqbal, Z. and Talegaonkar, S. 2016. Synergistic anticancer efficiency of bendamustine hydrochloride loaded bioactive hydroxyapatite nanoparticles: *In-vitro*, *ex-vivo* and *invivo* evaluation. Colloids Surf. B. Biointerface, 146:852-860.
- Tokatlian, T. and Segura, T. 2010. siRNA applications in nanomedicine. Wires Nanomed. Nanobi., 2:305-315.
- Turner, J.H. and Claringbold, P.G. 1991. A phase II study of treatment of painful multifocal skeletal metastases with single and repeated dose Samarium-153 ethylenediaminetetramethylene phosphonate. Eur. J. Cancer, 27(9):1084-1086.
- Turner, J.H., Claringbold, P.G., Hetherington, E.L., Sorby, P. and Martindale, A.A. 1989. A phase I study of Samarium-153 ethylene-diaminetetramethylene phosphonate therapy for disseminated skeletal metastases. J. Clinic. Oncol., 7(12):1926-1931.
- Turrens, J.F. 2003. Mitochondrial formation of reactive oxygen species. J. Physiol., 552(Pt 2):335-344.
- Uskokovic, V. and Uskokovic, D.P. 2011. Nanosized hydroxyapatite and other calcium phosphates: Chemistry of formation and application as drug and gene delivery agents. J. Biomed. Mater. Res., Part B, 96B(1):152-191.
- Venkatasubbu, G.D., Ramasamy, S., Avadhani, G.S., Ramakrishnan, V. and Kumar, J. 2013. Surface modification and paclitaxel drug delivery of folic acid modified polyethylene glycol functionalized hydroxyapatite nanoparticles. Powder Technol., 235:437-442.
- Venkatesan, P., Puvvada, N., Dash, R., Prashanth, K.B.N., Sarkar, D., Azab, B., Pathak, A., Kundu, S.C., Fisher, P.B. and Mandal, M. 2011. The potential of celecoxibloaded hydroxyapatite-chitosan nanocomposite for the

treatment of colon cancer. Biomater., 32(15):3794-3806.

- Vu, T.H., Shipley, J.M., Bergers, G., Berger, J.E., Helms, J.A., Hanahan, D., Shapiro, S.D., Senior, R.M. and Werb, Z. 1998. MMP-9/gelatinase B is a key regulator of growth plate angiogenesis and apoptosis of hypertrophic chondrocytes. Cell, 93(3):411-422.
- Wang, H., Zhai, L., Li, Y. and Shi, T. 2008. Preparation of irregular mesoporous hydroxyapatite. Mater. Res. Bull., 43:1607-1614.
- Weissig, V., Boddapati, S.V., Jabr, L. and D'Souza, G.G.
   2007. Mitochondria-specific nanotechnology.
   Nanomed. (Lond.), 2(3):275-285.
- Wuthier, R.E., Rice, G.S., Wallace, J.E., Weaver, R.L., Le Geros, R.Z. and Eanes, E.D. 1985. In vitro precipitation of calcium phosphate under intracellular conditions: Formation of brushite from an amorphous precursor in the absence of ATP. Calcif. Tissue Int., 37:401-410.
- Xia, Z., Liao, L. and Zhao, S. 2009. Synthesis of mesoporous hydroxyapatite using a modified hard-templating route. Mater. Res. Bull., 44:1626-1629.
- Xiong, H., Du, S., Ni, J., Zhou, J. and Yao, J. 2016. Mitochondria and nuclei dual-targeted heterogenous hydroxyapatite nanoparticles for enhancing therapeutic efficacy of doxorubicin. Biomater., 94:70-83.
- Xu, J., Xu, P.J., Li, Z.G., Huang, J. and Yang, Z. 2012. Oxidative stress and apoptosis induced by hydroxyapatite nanoparticles in C6 cells. J. Biomed. Mater. Res., Part A, 100A(3):738-745.
- Xu, Q., Tanaka, Y., Czernuszka, J.T. 2007. Encapsulation and release of a hydrophobic drug from hydroxyapatite coated liposomes. Biomat., 28:2687-2694.
- Yang, H.C. and Hon, M.H. 2009. The effect of molecular weight of chitosan nanoparticles and its application on drug delivery. Microchem, J., 92:87-91.
- Ye, F., Guo, H., Zhang, H. and He, X. 2010. Polymeric micelle-templated synthesis of hydroxyapatite hollow nanoparticles for a drug delivery system. Acta. Biomater., 6(6):2212-2218.
- Yuan, Y., Liu, C.S., Qian, J.C., Wang, J. and Zhang, Y. 2010. Size mediated cytotoxicity and apoptosis of hydroxyapatite nanoparticles in human hepatoma HepG2 cells. Biomat., 31(4):730-740.
- Zhang, C., Yang, J., Quan, Z., Yang, P., Li, C., Hou, Z. and Lin, J. 2009. Hydroxyapatite nano- and microcrystals with multiform morphologies: Controllable synthesis and luminescence properties. Cryst. Growth Des., 9:2725-2733.
- Zhao, F., Zhao, Y., Liu, Y., Chang, X., Chen, C. and Zhao, Y. 2011. Cellular uptake, intracellular trafficking, and cytotoxicity of nanomaterials. Small, 7(10):1322-1337.
- Zhivotovsky, B. and Orrenius, S. 2011. Calcium and cell death mechanisms: A perspective from the cell death community. Cell. Calcium, 50(3):211-221.
- Zhou, H. and Lee, J. 2011. Nanoscale hydroxyapatite particles for bone tissue engineering. Acta. Biomater., 7(7):2769-2781.
- Zhu, S.H., Huang, B.Y., Zhou, K.C., Huang, S.P., Liu, F., Li, Y.M., Xue, Z.G. and Long, Z.G. 2004. Hydroxyapatite nanoparticles as a novel gene carrier. J. Nanopart. Res., 6:307-311.

Zollen, M. 2011. CD44: Can a cancer-initiating cell profit from an abundantly expressed molecule? Nat. Rev. Cancer, 11:254-267. Zyman, Z.Z., Rokhmistrov, D.V. and Loza, K.I. 2013. Determination of the Ca/P ratio in calcium phosphates during the precipitation of hydroxyapatite using X-ray diffractometry. Proc. Appl. Ceram., 7:93-95.

## How to cite this article:

Ardhendu Kumar Mandal (2018) 'Hydroxyapatite Nanoparticles As Delivery System In Combating Various Diseases', *International Journal of Current Advanced Research*, 07(10), pp. 15869-15877. DOI: http://dx.doi.org/10.24327/ijcar.2018.15877.2911

\*\*\*\*\*\*