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RESEARCH ARTICLE

SALIVARY AMYLASE AND TOTAL PROTEIN LEVELS IN LEUKEMIAS

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ABSTRACT

Background: Oral manifestations are produced during different stages of leukemia and often reflects underlying systemic diseases, can be used as diagnostic indicator. This study was done to evaluate various oral complications produced by leukemia patients and to estimate the salivary amylase and salivary total protein levels in these patients to support the diagnostic value of saliva.

Material and Methods: A total of thirty patients of leukemia after confirmed diagnosis, who were not on chemotherapy drugs were included in the study group. 30 age and sex matched healthy individuals were enrolled as controls. Unstimulated saliva was analyzed for salivary amylase and salivary total proteins by autoanalyzer the same day.

Result: Patients in leukaemia group showed a significant increase in mean serum amylase levels when compared to controls ($p=0.02$). Although the mean salivary protein levels were higher in leukemia group but this difference was not significant statistically ($p=0.355$).

Conclusion: As leukemia patients showed higher salivary amylase and salivary total protein levels than control group and there is associated oral complications in leukemia group. This indicates decline in oral health with associated changes in saliva composition in leukemia patients. Hence supports the diagnostic value of saliva in coexisting systemic diseases such as leukemias.

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INTRODUCTION

Leukemias are malignant neoplasm of hematopoietic stem cells, characterized by diffuse replacement of the marrow by neoplastic cells. It is primarily a disease of the adults between the ages of 25 to 50 years with a peak incidence in fourth and fifth decades of life. They include: Lymphoid neoplasm – a diverse group of tumors of B-cell, T-cell and NK-cell origin, myeloid neoplasm, histiocytes: uncommon proliferative lesions of macrophages and dendritic cells.¹ Leukemias are also classified on the basis of clinical characteristics as acute and chronic leukemia.² Therefore categorized into acute myelogenous leukemia (AML), acute lymphocytic leukemia (ALL), chronic myelogenous leukemia (CML), and chronic lymphocytic leukemia (CLL).³

The etiology of leukemia is not well known and is considered as multifactorial. Risk factors associated with leukemia are smoking, radiation hazard, viral infections, chemical compounds like benzene, chromosomal abnormalities and/or family history of leukemia.⁴ Clinical features include fatigue, weakness, anorexia, weight loss, fever with or without identifiable infections, signs of abnormal hemostasis (bleeding, easy bruising), lymphadenopathy, nonspecific cough, headache, hepatomegaly, splenomegaly. Initial diagnosis of leukemia is mainly made on the basis of clinical presentation of the patient, followed by confirmed diagnosis based on identification of abnormal hematopoietic cells in blood and bone marrow aspirates. Further characterization is

done by cytochemical staining, immunophenotyping and cytogenetic analysis of chromosomal abnormalities.¹

As leukemia involves the whole body during different stages of disease progression resulting in a variety of oral manifestations, which are often mild and masked by the major manifestation of systemic disease.⁵ Among the different type of oral manifestation produced by leukemia, most frequent is paleness of oral mucosa/local abnormal color of the gum, gingival petechiae, ecchymosis, bleeding associating painless gingival hyperplasia, hemorrhages, ulcerative necrotic lesions and buccal infections.⁶

Saliva being diagnostic fluid offers advantages over blood/serum for its non-invasive nature, easy storage and transportation, cost-effectiveness and safe handling. These characteristics of saliva make it possible to monitor several biomarkers and also its use in many circumstances in which blood and urine sampling is not available. Saliva seems better for diagnostic purposes because of its linkage with traditional biochemical parameters which appears in different forms in the circulation.⁷ Hence this study was done to evaluate various oral complications produced by leukemia patients during their course of disease and to estimate salivary amylase and salivary total protein levels in these patients to support the diagnostic value of saliva.

MATERIALS AND METHODS

The present study was conducted in the Department of Biochemistry, in collaboration with the Department of

Medicine (Clinical Hematology unit); Pt. B.D. Sharma, University of Health Sciences, Rohtak. Thirty patients of leukemia after confirmed diagnosis, who were not on chemotherapy drugs were included in the study group. The diagnosis was made by history, clinical examination, total and differential leukocyte count, bone marrow examination and cytogenetic studies. 30 age and sex matched healthy volunteers were taken as controls.

Unstimulated saliva was obtained when subject seated quietly with his or her head flexed forward allowing the saliva to passively drip from the mouth to a collection container⁸ and then transferred to sterile collecting tube. Samples were centrifuged and pellet was discarded. Supernatant was analyzed for salivary amylase and salivary total protein by autoanalyzer the same day. IBM SPSS ver. 20 was used for various statistical analysis. Data was expressed in mean ± SD. P<0.05 was considered significant.

RESULT

In the present study thirty patients with confirmed diagnosis of leukemia were included from the department of Radiotherapy. Out of 30 cases, 11 (36.6%) had AML, 6 (20%) had ALL, 9 (30%) had CML and 4 (13.3%) had CLL. The age of study group varied from 17 to 68 years, with a mean age of 43.8 years. Out of 30 cases 22 were males and 8 were females. The AML group had 7 males and 4 females. The ALL group consisted of 4 males and 2 females, the CML group consisted of 7 males and 2 females and the CLL group had 4 males.

Table I Distribution of cases according to type of leukemias

Type of leukemia	No. of cases(out of 30)	Percentage (%)
AML	11	36.6%
ALL	6	20%
CML	9	30%
CLL	4	13.3%

Table II Distribution of cases according to Gender

Type of leukemia	Males	Females
AML	7	4
ALL	4	2
CML	7	2
CLL	4	Nil

Table III Comparison of salivary amylase and salivary total protein in controls and cases

Parameter	Control	Cases	Significance
Salivary amylase	28.45±11.60U/L	46.3±21.69U/L	P<0.05 Significant
Salivary total protein	0.38±0.10g/dL	0.41±0.10g/dL	p>0.05 Non-significant

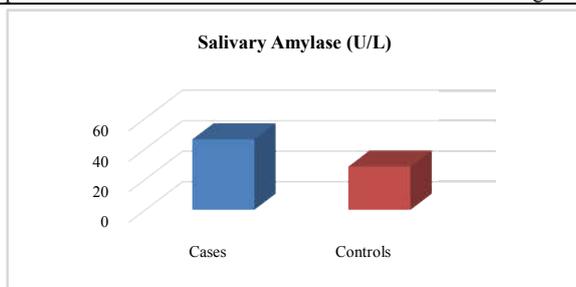


Figure I Salivary Amylase in control and cases

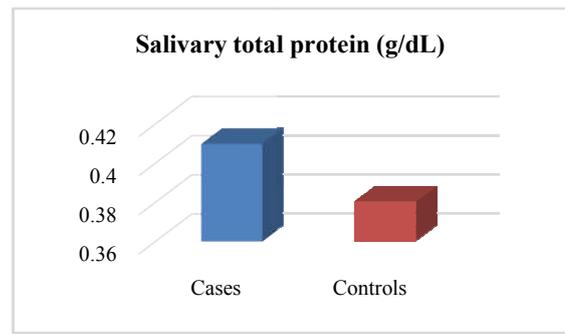


Figure II Salivary total proteins in control and cases

Table IV Comparison of salivary amylase and salivary total protein in acute and chronic leukemia

Parameter	Acute leukemia (n=17)	Chronic leukemia (n=13)	Significance
Salivary amylase	37.62±20.92U/L	59.75±13.53U/L	P<0.05 Significant
Salivary total protein	0.42±0.11g/dL	0.40±0.10g/dL	p>0.05 Non-significant

Salivary amylase in leukaemia patients ranged from 15-76U/L while in control group serum amylase value ranged from 14-58U/L. Mean serum amylase levels in leukemia patients were 46.3±21.69U/L and higher than the control group mean serum amylase levels being 28.45±11.60U/L and this difference was statistically significant (p=0.02). Mean serum amylase levels were 59.75±13.53U/L in chronic leukaemia patients and were significantly higher (p=0.016) than acute leukaemia patients salivary amylase levels being 37.62±20.92U/L.

The mean salivary total protein levels were 0.38±0.10g/dL in control group, while in leukaemia patient value was 0.41±0.10g/dL. This difference was not significant statistically (p= 0.355). In chronic leukaemia patients mean serum protein levels were 0.40±0.10g/dL and in acute leukemia value was 0.42±0.11g/dL. This difference was not significant statistically.(p=0.754)

Table V Oral manifestations in different leukemias

Oral manifestaion	AML (n=11)	ALL (n=6)	CML (n=9)	CLL (n=4)
Paleness of gum	9	5	7	2
Gingival hyperplasia	5	Nil	1	1
Gingival petechiae	3	2	Nil	Nil
Gingivitis	4	Nil	3	3
Enlarged submandibular lymphnodes	6	3	5	3
Ecchymosis	4	Nil	2	Nil
Ulcerative necrotic lesion	3	5	1	Nil

Out of 30 cases, 23 (76.6%) patients had discolouration (pallor) of the gum (9 AML, 5 ALL, 7 CML and 2 CLL patients); 7(23.3%) patients had gingival hyperplasia (5 AML, 1 CML and 1 ALL); 10 patients had gingivitis (4 AML, 3 CML and 3 CLL); 17 patient had enlarged submandibular lymph nodes (6 AML, 3 ALL, 5 CML and 3 CLL); 5 (16.7%) cases had gingival petechiae (3 AML and 2 ALL patients); and 6 (20%) patient had ecchymosis (4 AML, 2 CML); 9(30%) patients had ulcerative necrotic lesion (3 AML, 5 ALL and 1 CML) and none patient showed gum hemorrhages.

DISCUSSION

Leukemia during its different stages of progression involves the whole body system and produces systemic manifestations.

Oral manifestations often reflect the underlying systemic disease and therefore saliva has been used recently as a diagnostic indicator of oral health⁹⁻¹¹ in leukemia and many other systemic diseases where function of salivary gland and composition of saliva is affected. Salivary components are now used as surrogates for systemic biomarkers in blood.¹² Our study reported mean salivary total protein levels 0.38 ± 0.10 g/dL in control group, while the mean values in leukemia patient was 0.41 ± 0.10 g/dL. Although salivary protein levels are higher in leukemic patients but this difference was not significant statistically ($p = 0.355$) (Table III, Figure II).

In the oral cavity saliva serves as the first barrier defending against the microbial invasion by mechanical, nonimmunologic (non-specific) and immunologic (specific) means.¹³ Salivary proteins (e.g. lysozyme, peroxidase, myeloperoxidase, lactoferrin) along with salivary components (thiocyanate, chlorine, hydrogen peroxide) are mainly responsible for innate immunity via their immune activator and/or immune modulator properties.^{13,14} Their levels undergo considerable fluctuations in various physiological and pathological conditions such as physical exercise, age, tobacco smoke, diabetes, hyper-IgE syndrome and caries etc.¹⁵

The effect of these salivary proteins are cumulative and/or synergistic providing molecular defense in the oral cavity. Local concentration of these proteins near the mucosal surfaces (mucosal transudate), periodontal sulcus (gingival circular fluid) and oral ulcers (transudate) is very high and also reinforced by immune and/or inflammatory reactions of oral mucosa. Binding of salivary proteins onto mucosal surfaces improves their resistance against proteolytic degradation via microbial proteases.¹⁴ They interfere with bacterial metabolism, its adhesion and cause bacterial aggregation, so that they can be swallowed.¹³ The salivary levels of total protein increase through β -sympathetic activity in the salivary glands, since salivary secretion is mainly evoked by the action of adrenergic mediators.¹⁶ Salivary glands may respond to periodontitis by enhanced synthesis of some acinar proteins and thereby an increase in the protective potential of saliva.¹⁷⁻¹⁹

The mean serum amylase levels in our study in leukemia patients were 46.3 ± 21.69 U/L and higher than the control group mean serum amylase levels being 28.45 ± 11.60 U/L and this difference was statistically significant ($p = 0.02$) (Table: III, Figure: I).

Amylase is a highly abundant protein in saliva, having endoglycosidase activity. It has a role in acquired pellicle formation on tooth surfaces. In salivary fluid amylase is one of the most important digestive enzymes and thus α -amylase is prone to alterations in response to the cell damage caused by chronic periodontitis. α -amylase has a higher concentration in saliva and are main constituents of the glycoproteinaceous thin film formed on teeth. Formation of a preliminary film is later followed by the dental plaque formation which contributes to various oral diseases, dental caries and periodontitis.^{20,21} The salivary α -amylase can bind specifically to the most common oral bacteria, streptococcus species.²²

Amylase inhibits bacterial growth by binding to lipopolysaccharide of bacteria which is a structural part of

bacteria. It also inhibits bacterial toxin-mediated tissue destructive inflammatory reactions.²³ Thus, it was demonstrated that α -amylases from human saliva, porcine pancreas and rice show a significant cell growth inhibitory activity against *Porphyromonas gingivalis* species, and interfere with the adherence and biofilm formation of *Aggregatibacter actinomycetemcomitans*, indicating that α -amylase could be effective in preventing periodontal diseases.^{24,25}

Unstimulated saliva was used because there is a prevalent resting condition in the oral cavity for most of the 24-hour period and, as the salivary secretion is a reflex response which can be influenced by several stimuli such as the accumulation of plaque-derived substances or inflammatory products.²⁶ Other studies had reported a positive relationship between basal values of amylase, proteins and mucin with CAL (clinical attachment loss) and PPD (probing pocket depth).²⁷ This is mainly explained by the fact that inflammatory products may trigger salivary secretion via neural pathways.²⁶

Levels of amylase, mucin and proteins in saliva from adults with or without chronic periodontal disease were evaluated in a previous study. They reported higher concentrations of mucin, amylase and proteins were found in saliva from patients compared with healthy subjects.¹⁷ Ashok L *et al* also reported raised salivary amylase and salivary protein levels in leukemia patients when compared with healthy controls.²⁸

Out of 30 cases, 23 (76.6%) patients had discoloration (pallor) of the gum (9 AML, 5 ALL, 7 CML and 2 CLL patients); 7 (23.3%) patients had gingival hyperplasia (5 AML, 1 CML and 1 ALL); 10 patients had gingivitis (4 AML, 3 CML and 3 CLL); 17 patients had enlarged submandibular lymph nodes (6 AML, 3 ALL, 5 CML and 3 CLL); 5 (16.7%) cases had gingival petechiae (3 AML and 2 ALL patients); and 6 (20%) patients had ecchymosis (4 AML, 2 CML); 9 (30%) patients had ulcerative necrotic lesions (3 AML, 5 ALL and 1 CML) and none of the patients showed gum hemorrhages (Table: V).

Acute leukemia patients often present to dentists with different types of oral manifestations at various stages of disease. Gingival enlargement is the most commonly seen oral manifestation in leukemia patients. Other manifestations include paleness of oral mucosa/local abnormal color of the gum, gingival petechiae, ecchymosis, hemorrhages, ulcerative necrotic lesions and buccal infections.⁶ In chronic leukemia oral manifestations are non-specific and less common. In these patients oral lesions occur because of infiltration in local tissues by leukemic cells, anemia, thrombocytopenia and immunodeficiency state. Dreizen *et al* also reported gingival hyperplasia in acute leukemia patients both ALL and AML, which disappears totally or partially in most of the cases after chemotherapy administration.²⁹⁻³¹

CONCLUSION

As leukemia patients showed higher salivary amylase and salivary total protein levels than control group and there are associated oral complications in leukemia group. This indicates a decline in oral health with associated changes in saliva composition in leukemia patients. Hence, it supports the diagnostic value of saliva in coexisting systemic diseases such as leukemias.

References

1. Aster CJ. Diseases of white blood cells, lymph nodes, spleen and thymus. In: Kumar V, Abbas AK, Fausto N, Atser JC editors. Robbins and Cotran pathological basis of disease. 7thed New York: Elsevier 2004;661-710.
2. Haouas H, Haouas S, Uzan G, Hafsia A. Identification of new markers discriminating between myeloid and lymphoid acute leukemia. *Hematology* 2010;15:193-203.
3. Wu J, Fantasia JE, Kaplan R. Oral manifestations of acute myelomonocytic leukemia: a case report and review of the classification of the leukemias. *J periodontol* 2002;73:664-8.
4. Rashid A, Lee NG, Jakobiec FA, Freitag SK. Epstein-Barr virus-positive polymorphous lymphoblastic infiltrate of the lacrimal glands in a patient with acute lymphoblastic leukemia. *JAMA Ophthalmol* 2014;132:892-4.
5. Henderson ES, Lister TA. Leukemia. 5th ed. Philadelphia: W. B. Saunders Company: 1990.
6. Deliverska EG, Krasteva A. Oral signs of leukemia and dental management- literature data and case report. *IMAB* 2013;19:388-91.
7. Kaufman E, Lamster IB. The diagnostic applications of saliva- A review. *Crit Rev Oral Biol Med* 2002;13:197-212.
8. Munro CL, Grap MJ, Jablonski R, Boyle A. Oral health measurement in nursing research: state of science. *Biological Research for Nursing* 2006;8: 35-42.
9. Lee A, Ghaname CB, Brauen TM. Bacterial and salivary biomarkers predict the gingival inflammatory profile. *Journal of Periodontology* 2012;83:79-89.
10. Sexton WM, Lin Y, Kryscio RJ, Dawson DR, Ebersole JL, Miller CS. Salivary biomarkers of periodontal disease in response to treatment. *Journal of Clinical Periodontology* 2011;38: 431-41.
11. Kang EH, Lee YJ, Hyon JY, Yun PY, Songg YW. Salivary cytokine profiles in primary sjogren's syndrome differ from those in non-sjogren sicca in terms of TNF- α levels and Th-1/Th-2 ratios. *Clinical and Experimental Rheumatology* 2011;29: 970-6.
12. Miller CS, Foley JD, Bailey AL, Campell CL, Humphries RL, Christodoulides N, *et al.* Current developments in salivary diagnostics. *Biomark Med* 2010;4: 171-89.
13. Karolewska E, Konoka T, Pupek M, Chaber R. Mucositis in children with leukemia and salivary defense factors. *Dent Med Probl* 2007;44:30-6.
14. Fabian TK, Hermann P, Beck A, Fabian G. Salivary defense proteins: their network and role in innate and acquired oral immunity. *Int J MolSci* 2012;13: 4295-320.
15. Koscielniak D, Jurczak A, Zygmunt A, Krzysciak W. Salivary proteins in health and disease. *Acta Biochimica Polonica* 2012; 59:451-7.
16. Chicharro JL, Lucia A, Perez M, Vaquero AF, Urena R. Saliva composition and exercise. *Sports Med* 1998;26:17-27.
17. Sanchez GA, Miozza VA, Delgado A, Bush L. Determination of salivary levels of mucin and amylase in chronic periodontitis patients. *J Periodontal Res* 2011; 46:221-7.
18. Kejriwal S, Bhandary R, Thomas B, Kumari S. Estimation of levels of salivary mucin, amylase and total protein in gingivitis and chronic periodontitis patients. *J Clin Diag Res* 2014; 8:ZC56-60.
19. Henskens YM, van den Keijbus PA, Veerman EC, Van der Weijden GA, Timmerman MF, Snoek CM, *et al.* Protein composition of whole and parotid saliva in healthy and periodontitis subjects. Determination of cystatins, albumin, amylase and IgA. *J Periodontal Res* 1996; 31:57-65.
20. Chaudhuri J, Vikerman M, Tanzer JM, Scannapieco FA. Interaction of salivary alpha-amylase and amylase-binding protein A (AbpA) of streptococcus gordonii with glycosyl transferases of S. gordonii and streptococcus mutans. *BMC Microbiology* 2006; 7:60-9.
21. Paknjad M, Rezaei A. Salivary biochemical markers of periodontitis. *Rom J Biochem* 2013; 50:129-46.
22. Vashist N, Kiran AR, Vashist D. Gingival enlargement leading to the diagnosis of acute lymphoblastic leukemia in an 8-year old girl: A case report. *Journal of Dental and Medical Sciences* 2013; 7:67-70.
23. Fabian TK, Hermann P, Beck A, Frejerdy P, Fabian G. Salivary defense proteins: their network and role in innate and acquired oral immunity. *Int J Mol Sci* 2012; 13:4295-320.
24. Baik JE, Hong SW, Choi S, Jeon JH, Park OJ, Cho K, *et al.* Alpha-amylase is a human salivary protein with affinity to lipopolysaccharide of *Aggregatibacter actinomycetemcomitans*. *Mol Oral Microbiol* 2013;28:142-53.
25. Ochiai A, Harada K, Hashimoto K, Shibata K, Ishiyama Y, Mitsui T, *et al.* α -Amylase is a potential growth inhibitors of porphyromonas gingivalis, a periodontal pathogenic bacterium. *J Peridont Res* 2014; 49:62-8.
26. Seeman R, Hagewald SJ, Sztankay V, Drews J, Bizhang M, Kage A. Levels of parotid and submandibular/sublingual salivary immunoglobulin A in response to experimental gingivitis in humans. *Clin Oral Invest* 2004; 8:233-7.
27. Sanchez GA, Miozza VA, Delgado A, Bush L. Relationship between salivary mucin or amylase and the periodontal status. *Oral Dis* 2013;19:585-91.
28. Ashok L, Sujatha GP, Hema G. Estimation of salivary amylase and total proteins in leukemia patients and its correlation with clinical feature and radiographic finding. *Indian J Dent Res* 2010;21:486-90.
29. Dreizen S, McCredie KB, Keating MJ, Luna MA. Malignant gingival and skin "infiltrates" in adult leukemia. *Oral Surg Oral Med Pathol* 1983; 55: 572-9.
30. Curtis I, Cooper CL, Loewen R, Shore T. Gingival hyperplasia complicating acute myelomonocytic leukemia. *J Can Dent Assoc* 2000; 66:78-9
31. Beltzer EK, Fortunato CK, Guaderrama MM, Perkins MK, Garramone BM, Granger DA. Salivary flow and alpha-amylase: Collection technique, duration, and oral fluid type. *Physiology and Behaviour* 2010;2: 289-96.