



PHENOTYPIC PROFILING OF VIRULENT ESCHERICHIA COLI ISOLATED FROM URINARY TRACT INFECTIONS FROM WESTERN DISTRICTS OF TAMIL NADU

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ABSTRACT

Objective: Urinary tract infection is a frequently encountered problem in developing and developed nations. There are various methods to identify UTI pathogens with different specificity and sensitivity. The study focuses on monitoring the prevalence of *E.coli* in patients with Urinary tract infections and UTI suspected patients.

Methods: The methods include preliminary processing of samples by culturing in BHI and screening of *E.coli* in EMB Agar. Further microbiological and biochemical characterization were carried out for identification.

Results: A predominant presence of pathogenic *E.coli* accounting for more than 50% of UTI was observed.

Conclusion: The results are indicative of an efficient screening and isolation method for *E.coli* in patients with UTI and UTI suspected patients. Moreover the antibiotic sensitivity screening patterns will aid in the treatment regime of the cases

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INTRODUCTION

Escherichia coli is a common inhabitant of the gastrointestinal tract of humans and animals. Usually, *E. coli* forms a beneficial symbiotic relationship with its host and plays important roles in promoting the stability of the luminal microbial flora and in maintaining normal intestinal homeostasis (Yan and Polk, 2004). As a commensal, *E. coli* rather remains harmlessly confined to the intestinal lumen and rarely causes a disease. However, in the debilitated or immuno suppressed host, or when the gastrointestinal barriers are violated, even nonpathogenic-commensal strains of *E. coli* can cause infection (Kaper et al., 2004). Some strains of *E. coli* can diverge from their commensal cohorts, taking on a more pathogenic nature. These strains acquire specific virulence factors (via, DNA horizontal transfer of transposons, plasmids, bacteriophages, and pathogenicity islands) which confer an increased ability to adapt to new niches and allow the bacteria to increase the ability to cause a broad spectrum of diseases. Successful establishment of infection by bacterial pathogens requires adhesion to host cells, colonization of tissues, and, in certain cases, cellular invasion, followed by intracellular multiplication, dissemination to other tissues or persistence. Colonization of the urine in the absence of the clinical symptoms is called asymptomatic bacteriuria (ABU).

Most patients with ABU do not need treatment, and in many cases the colonizing by the ABU strains may help to prevent infection by other more virulent bacteria (Hull et al., 2000; Darouichee et al., 2001; Trautner et al., 2002). The primary causative agents responsible for more than 80% of all UTIs, including both ABU and symptomatic UTIs, are strains of uropathogenic *E. coli* (Hooton and Stamm, 1997; Svanborg and Godaly, 1997). Antibiotic treatment, typically with trimethoprim / sulfamethoxazole or ciprofloxacin, is generally effective for eradication of the infecting strain.

The current study is aimed at the incidence of UPEC despite previous studies since reports on UPEC in children are not available in this part of the state. UPEC strains encode a number of virulence factors, which enable the bacteria to colonize the urinary tract and persist in face of highly effective host defense. UPEC isolates exhibit a high degree of genetic diversity due to the possession of specialized virulence genes located on mobile genetic elements called pathogenicity islands (Oelschlaeger et al., 2002; Wiles et al., 2008). Attention has been given on the multiple antibiotic resistant and haemolysin positive strains of UPEC in clinical and diarrhoeal samples. In addition, the strains have been screened and characterized.

MATERIALS AND METHODS

Bacterial strain and isolates: A total of 137 urine samples (from patients having UTI and suspected UTI individuals) were collected from various hospital-attached laboratories and

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private clinical laboratories located various places from the Districts of Coimbatore, Erode, Tiruppur, The Nilgiris and Salem. Collection of samples was done during the period from November 2014 to May, 2015. Throughout the study the standard strain of *E. coli* MTCC 326 was used as the positive control.

Growth and screening of UTI *E. coli*: All the urine samples were enriched in Brain heart infusion broth (BHIB) (HiMedia, India). 0.1 ml each samples were transferred to 99 ml of sterile BHI broth medium and incubated for 18 hrs at 37 °C. The resulting colonies were further streaked onto Eosine Methylene Blue (EMB) agar (HiMedia, India) and the samples showing green metallic sheen after 24 h of incubation were primarily identified as *E. coli*. Additionally, the isolates were analyzed for lactose fermentation by plating on MacConkey Agar (HiMedia, India).

Microbiological and Biochemical characterization: The green metallic colonies of *E. coli* were confirmed by Gram staining. Biochemical tests were performed according to Bergey’s Manual of Determinative Bacteriology (Ref).

Antibiotic sensitivity test and MAR indexing: The antibiotic sensitivity test was done on Mueller-Hinton agar by Kirby-Bauer disc diffusion test as per Clinical and Laboratory Standard Institute (CLSI) guidelines. The isolates were tested for ampicillin (10 µg), cefuroxime (30 µg), ceftriaxone (30 µg), norfloxacin (10 µg), nitrofurantoin (300 µg), amoxicillin-clavulanic acid (10/20 µg), co-trimoxazole (1.25/23.75 µg), cefepime (30 µg), ciprofloxacin (5 µg), amikacin (30 µg), piperacillin-tazobactam (100/10 µg) and imipenem (10 µg) (Hi-media, Mumbai). An isolate was considered as MDR if found resistant to three or more antimicrobials belonging to different classes/groups of antimicrobials. Multiple antimicrobial resistance (MAR) profiles were also examined using β-lactam (17) and non β-lactam (42) antimicrobial agents. Bactericidal activity (serum susceptibility) of *E. coli* on normal human serum was also calculated.

RESULTS AND DISCUSSION

Bacterial strain and isolates

Table 1 Samples, sampling area and sample data

Month	Sampling area	No of urine samples		Total	Total
		UTI samples	Non-UTI samples		
Nov-14	Coimbatore	6	1	7	22
	Erode	2	0	2	
	Tiruppur	0	1	1	
	Kangeyam	1	0	1	
	Annur	3	2	5	
	Salem	2	2	4	
	Ooty/Coonoor	1	1	2	
	Coimbatore	2	1	3	
	Erode	3	1	4	
Dec-14	Tiruppur	1	0	1	20
	Kangeyam	-	-	0	
	Annur	3	1	4	
	Salem	4	2	6	
	Ooty/Coonoor	2	0	2	
Jan-14	Coimbatore	2	0	2	19
	Erode	3	1	4	
	Tiruppur	2	1	3	

Feb-14	Kangeyam	--	--	--	27
	Annur	2	3	5	
	Salem	3	0	3	
	Ooty/Coonoor	2	0	2	
	Coimbatore	15	-	15	
	Erode	2	1	3	
	Tiruppur	6	0	6	
	Kangeyam	3	0	1	
	Annur	1	1	2	
Mar-14	Salem	--	--	--	24
	Ooty/Coonoor	--	--	--	
	Coimbatore	4	1	5	
	Erode	1	2	3	
	Tiruppur	3	0	3	
	Kangeyam	4	1	5	
	Annur	-	1	1	
	Salem	2	2	4	
	Ooty/Coonoor	3	0	3	
Apr-14	Coimbatore	1	-	1	12
	Erode	0	0	0	
	Tiruppur	4	0	4	
	Kangeyam	2	1	3	
	Annur	1	1	2	
	Salem	0	1	1	
	Ooty/Coonoor	1	0	1	
	Coimbatore	2	1	3	
	Erode	3	1	4	
May-14	Tiruppur	1	2	3	13
	Kangeyam	3	0	3	
	Annur	0	0	0	
	Salem	0	0	0	
	Ooty/Coonoor	0	0	0	
	Total	106	33	137	

All the specimens were labeled as UTE 01 to UTE 106; NUT 01 to 33

Growth and screening of UTI *E. coli*

Total No. of samples	Positive Green metallic sheen colony			
	UTI samples (n=106)	% of incidence	Non UTI samples (n=33)	% of incidence
137	57	53.77	8	24.3

Percentage incidence of LF and NLF bacterial growth on MacConkey agar

No. of Samples Collected	UTI samples		Non UTI Samples	
	LFB*+ve	%	LFB*+ve	%
137	59	55.66	16	48.48

LFB* - Lactose fermenting bacteria; NLFB** - Non Lactose fermenting bacteria

CONCLUSION

The result of the present study shows a prevalence and distribution of *E.coli* in patients with UTI and suspected UTI. Further studies on the virulence and pathogenicity of the strains are proposed to be carried out to better understand the role of pathogenic *E.coli* in the disease pathogenesis of UTI, since pathogens related to these infections are gaining importance due to sudden outbreaks.

Microbiological and Biochemical characterization

Biochemical Tests	<i>E. coli</i>		UTE																			
	MTCC 326	Metallic sheen	1	2	3	4	6	7	8	10	11	13	14	16	18	19	20	24	25	26	29	30
Colour (on EMB)	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Gram-stain	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Cytochrome oxidase	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Catalase	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Motility	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
O/F	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Methyl red	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
VP	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
H ₂ S	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Indole	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Citrate	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
ONPG	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Nitrate-Nitrite	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Acid (-A) and Gas (-G) from:																						
Arabinose	A	A	A	A	A	AG	A	A	A	A	A	AG	A	A	A	AG	A	A	A	A	A	A
Cellobiose	AG	AG	AG	AG	AG	A	AG															
Fructose	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Galactose	A--	A--	A--	AG	A--																	
Glucose	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	A-	AG									
Maltose	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A	A
Mannitol	--	--	--	--	--	--	--	A	--	--	--	G	--	--	--	--	--	--	--	G	--	--
Mannose	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG
Sorbitol	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG	AG

Antibiotic sensitivity test and MAR indexing: The antibiotic sensitivity testing revealed normal sensitivity and some resistant patterns. The results were arrived with the comparison of MTCC 326 as a standard.

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