



## **EFFICACY OF CITRIC ACID AND CHLORHEXIDINE AS DENTURE CLEANSERS AGAINST CANDIDA ALBICANS – AN IN VITRO STUDY**

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### **INTRODUCTION**

A complete denture is defined as *a dental prosthesis, which replaces the entire dentition and associated structures of the maxilla and mandible* (1). A denture becomes susceptible to infections due to the biofilm formed on its surface on placement in the oral environment (2). Almost 11-67% of the denture wearers have reported with Denture stomatitis (3). *Candida* is one of the chief causes for denture-induced stomatitis due to its adherence to denture surface and oral tissues, its ability to form biofilms and resistance to antifungal agents (4). Decreased flow of oxygen and saliva caused by the denture results in local acidic and anaerobic microenvironment of the underlying tissues that favours yeast growth. This ultimately increases the prevalence of *Candida* by 60-100% in denture wearers (5). *Candida albicans* that manifest in the biofilm, an important contributor to the pathogenesis of denture stomatitis, is essential for the instalment and maintenance of denture stomatitis (6).

Dentures can be cleaned mechanically, chemically or through a combination of both these methods. Mechanical methods comprises of brushing, and ultrasonic treatment uses ultrasonic cleansers (7). Chemical methods include soaking the dentures in commercial (peroxides, acids, mouth washes and enzymes) or household (hypochlorides, sodium chloride vinegar) products (8). These chemicals are easy to use and can easily reach undercuts of the denture base which are otherwise overlooked during other denture cleansing methods. One of the main disadvantages of mechanical denture cleansing methods is abrasion due to brushing is overcome by chemical methods as the acrylic resins surface roughness remains unchanged and the surfaces are less susceptible to biofilm accumulation (9).

Citric acid denture cleanser is available as a concentrated solution, which can be used 1:5 dilution daily or 1:8 dilution weekly after proper dilution (as indicated by the manufacturer).

This cleanser acts as a chemotherapeutic agent that can effectively disrupt biofilms through a sequestering mechanism with calcium ions. This mechanism allows citric acid to break calcium bridges and subsequently disrupt the biofilm matrix, which may lead to anti-biofilm activity. Although citric acid cleansers used continuously for 3 months showed adverse effects with greater ion release from Co–Cr alloys when used to decontaminate implant surfaces, no harmful effects have been demonstrated with denture materials in general. Therefore, citric acid cleansers might be suitable for removable dentures and orthodontic appliances and for removing biofilms and preventing their recolonization (10).

Chlorhexidine has a broad spectrum of antimicrobial activity for a number of organisms including *Candida* and is used as a topical therapeutic supplement (11, 12). Also candidal adhesion to biological and inert surfaces can be inhibited by chlorhexidine (13). Chlorhexidine acts as a fungicide and has fungistatic action which results in coagulation of nucleoproteins and escape of cytoplasmic components through the plasmolemma due to cell wall changes (14).

There are many known denture disinfectants such as EDTA, sodium hypochlorite, sodium perborate, povidone iodine, hydrogen peroxide, etc. (15). In this study we study the effectiveness of citric acid and chlorhexidine as denture cleansers.

### **Methodology**

The effect of disinfectant was tested by two methods. One was by contamination of denture bases with *Candida* suspension and the second method was by testing the effect of the standardized concentration of disinfectant in a broth.

### **Sample fabrication**

A total of 40 heat-polymerized acrylic denture strips were obtained from a wax pattern with a standardized dimension of 5x1cm. The wax pattern was invested with dental stone (type III gypsum) in a metallic flask. After the setting of dental stone, dewaxing is done by immersing the flask in a water bath at a temperature of 70-80°C for about 10 minutes (18). Heat-polymerized acrylic resin was mixed according to the manufacturers recommendation and packed into the mold at the dough stage. The metal flask was then closed and subjected to a short curing cycle at 74°C for 2 hours followed

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by a terminal boiling at 100°C for 1 hour (19). On completion of curing cycle, the flask was allowed to completely cool before opening and the denture sample was obtained. The denture strips of 5x1cm dimension were checked for any imperfections. The cameo surface of the strips were sandpapered and polished (19). On completion of processing, the strips were packed and autoclaved.

## Contamination of specimen

40 heat cured denture acrylic denture strips were selected and sterilized by autoclaving at 15lbs for 30 minutes. These denture strips were immersed in sterilized uricol containers containing 50ml of sterilized artificial saliva. A Candida albicans suspension was made to the turbidity matching 0.5 Mcfarland standard by immersing for 30 minutes. 100µl of suspension is added to the artificial saliva and well shaken to ensure a good mix. The denture strips in the above suspension was incubated for 3 days at 37° Celsius after which it was taken out, and cleaned with mineral water and then immersed in 50ml of disinfectant and kept for 6 hours. A subculture was made on Brain Heart Infusion agar and incubated for 24 hours.

## Preparation of disinfectants

Commercially available 20% citric acid and 2% chlorhexidine were used as denture cleansing agents in this study. Saline was taken as the negative control and 0.2% chlorhexidine containing commercially available mouthwash was taken as the positive control. After incubation for 48 hours, the denture samples were washed in drinking water and placed in a sterile container containing denture cleansing agent. 10 denture samples were placed in each denture cleansing agent (20% citric acid and chlorhexidine 2%). The denture samples were left in the denture cleansing agent for 6 hours.

## Culture preparation

After 6 hours, a swab was taken from the rough surface of the denture base sample and streak on the SDA plate. Repeat this for all the denture base samples. Incubate the SDA plates for 24 hours. After 24 hours, the growth pattern of Candida albicans was observed.

## Broth culture

The disinfectant material is taken in a standardized concentration in 5 cuvettes of 1ml each, the candida suspension which was made with turbidity matching 0.5 McFarland standard is taken and 10 microliter of the suspension is added to disinfectants taken in cuvette. It was allowed to react for 6 hours at room temperature. After the 6 hour period 10 microliter of this preparation was transferred to saborauds dextrose agar and incubated for 12 hours at 37 degrees Celsius. The test was done along with a positive and a negative control.

## Experimental and control groups

Four groups each containing contaminated specimen of 10 were assigned to various disinfectants.

- Group 1: Saline (control)
- Group 2: Citric acid 20%
- Group 3: Chlorhexidine 0.2%

Group 4: Chlorhexidine 2%

Table 1

Denture Cleansing agent	Positive	Negative
Citric acid 20%	3	7
Chlorhexidine 2%	0	10
Saline	10	0
Chlorhexidine 0.2%	4	6

## RESULTS

The results obtained from both methods were consistent with each other. When tested against the two controls, it was found that both disinfectants had effective disinfecting properties against Candida albicans.

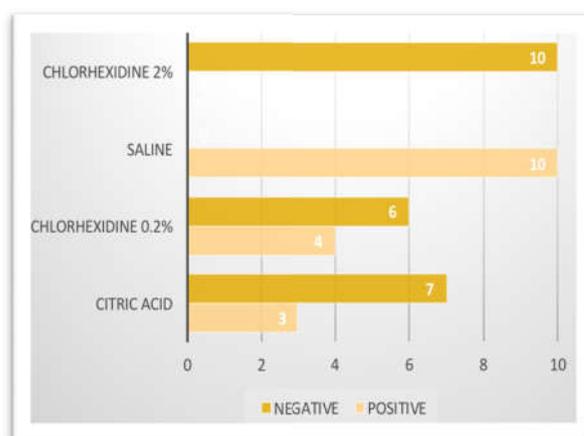
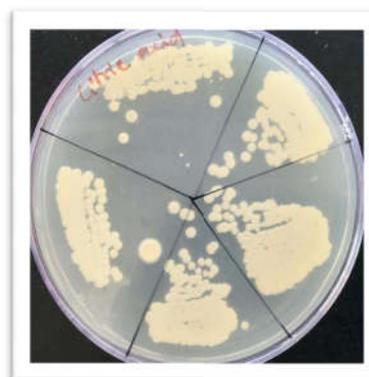
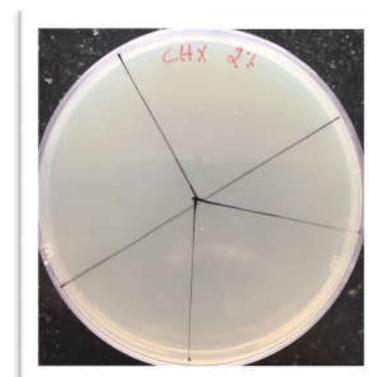


Fig 1



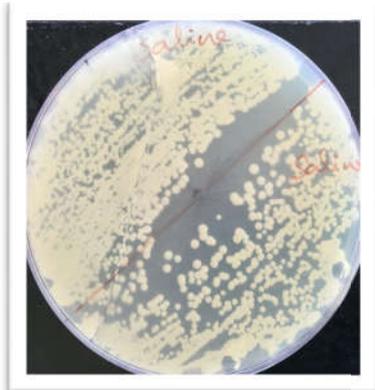
Citric acid



Chlorhexidine 2%



Chlorhexidine 0.2%



Saline

## DISCUSSION

Citric acid is a chemotherapeutic agent with an increased potential for eliminating biofilms *in vitro*, although complete removal of the biofilms is not reported (16,17). The current study revealed a 40% positive result for *Candida* when 20% citric acid was used as a denture cleanser. One study showed that although the number of viable cells immediately after treating with citric acid was null, it was confirmed that a residual biofilm was present (10). Citric acid has showed reduction in the viability of *Candida albicans* immediately after treatment but cannot completely cleanse the denture surface or prevent the recolonisation of the candidial biofilm (10). Citric acid does not affect the basal layers of the biofilm which is protected by an extra cellular matrix and hence leads to the recolonisation of *Candida* after 48 hours. Citric acid will thus not be effective in eliminating a biofilm of *Candida* or prevent their recolonisation on a single exposure (18, 19). Citric acid acts by disrupting the biofilms but it does not completely remove them. This finding suggests that citric acid can be an effective complementary method for biofilm removal after the debris are removed from the denture surface namely by using a mechanical method. Thus, citric acid is expected to present more consistent effects on biofilm elimination when used as an adjunct to mechanical methods, such as brushing, to completely cleanse the dentures (16,17,20).

*Candida* species might express increased virulence in addition to adherence in biofilms formed and hence are more resistant to antifungals (21, 22). Studies showed that microorganisms present in the biofilm have an increase in resistance to antimicrobial agents up to 500 times that of planktonic organisms (23, 24). Hence it is very essential to employ new treatment options for *Candidiasis* though chlorhexidine is not

the first line of treatment against *Candida* (25). Chlorhexidine acts by binding to the negatively charged surfaces like the epithelial cells because it is a cationic biguanide (25) and it disrupts the adhesion abilities of *Candida* which is an important virulent factor (13). Chlorhexidine's effect at a macroscopic level shows disruption of adherence and at cellular level indicates the effect of chlorhexidine on the structural viability and integrity of *Candida* biofilms (26). At low doses chlorhexidine damages the cellular transport of bacterial cells by creating pores in the cell membrane and at higher doses will cause cell death by penetrating into the microbial cell (27). Studies prove this destruction capacity of chlorhexidine where the number of viable cells in the *Candida* biofilm were significantly reduced after exposure to chlorhexidine (25). The current study tested chlorhexidine of 2 different concentrations where chlorhexidine 0.2% had 4 strips that were positive for *Candida* whereas the chlorhexidine 2% had no strips positive for *Candida* (Table 1.1) indicating the effectiveness of Chlorhexidine against *Candida* biofilms at higher concentrations.

In the present study citric acid 20% had a success rate of 70% in eliminating *Candida* biofilms. Previous studies conducted had similar results where complete removal of *Candida* was seen as an immediate effect to citric acid exposure. However these studies indicated the recolonisation of *Candida* in 48 hours (10). This shows that citric acid has to be used as a complementary agent in denture cleansing as it can remove biofilms though not permanently. Chlorhexidine that was tested in the present study revealed results based on the concentration where chlorhexidine 0.2% had a success rate of 60%, chlorhexidine 2% had a success rate of 100% as all the strips were negative for *Candida*. This is in accordance with studies that showed the different mechanisms of action of varying chlorhexidine concentrations (27). Hence from this study we can conclude that both the disinfectants are effective in eliminating *Candida* biofilms. Citric acid can be used either in combination with mechanical methods of brushing or coupled with another chemical disinfectant that can complement its properties. Similarly chlorhexidine can be included in a comparatively higher concentration in denture cleansers to prevent *Candidal* manifestations as *Candida* have developed resistance to the commonly used antifungals. However large scale studies regarding the efficacy of such methods is required to establish a reliable result.

## CONCLUSION

Hence of the two chemical disinfectants studied, three strips showed positive candidial growth for Hydrogen peroxide and none of the strips showed positive candidial growth for Chlorhexidine 2% concluding that both agents are effective denture cleansers.

## References

1. Sharry JJ. Complete Denture Prosthodontics, 3rd ed. New York: Blakiston; 1974: 191-210.
2. The Health and Social Care Information Centre. Adult dental health survey 2009. London: Department of Health, 2010.
3. Theilade E, Budtz-Jorgensen E. Predominant cultivable microflora of plaque on removable dentures

- in patients with denture induced stomatitis. *Oral Microbiol Immunol* 1988; 3:8-13
4. Silva M, Vergani C, Giampaolo E, Neppelenbroek K, Spolidorio D, Machado A. Effectiveness of microwave irradiation on the disinfection of complete dentures. *Int J Prosthodont* 2006; 19:288-293
  5. Fungal biofilms and related infections advances in Microbiology, Infectious Diseases and Public Health Volume 3 page 89
  6. Dorey JL, Blasberg B, MacEntee MI, Conklin RJ. Oral mucosal disorders in denture wearers. *JProsthetDent*. 1985;53(2):210-3
  7. Sanita P, Vergani C, Giampaolo E, Pavarina A, Machado A. Growth of Candida species on complete dentures: effect of microwave disinfection. *Mycoses* 2009; 52:154-160
  8. Senna P, Sotto-Maior B, Silva W, Del Bel Cury A. Adding denture cleanser to microwave disinfection regimen to reduce the irradiation time and the exposure of dentures to high temperatures. *Gerodontology* 2013; 30:26-31
  9. Senna P, da Silva W, Cury A. Denture disinfection by microwave energy: influence of Candida albicans biofilm. *Gerodontology* 2012; 29:186-191
  10. Fernanda Faot, Yuri WanderleyCavalcanti, Martinna de Mendonça e Bertolini, Luciana de Rezende Pinto, Wander José da Silva, Altair Antoninha Del Bel Cury. Efficacy of citric acid denture cleanser on the Candida albicans biofilm formed on poly (methyl methacrylate): effects on residual biofilm and recolonization process. *BMC Oral Health* 2014, 14:77
  11. Suci PA, Tyler BJ. Action of chlorhexidine digluconate against yeast and filamentous forms in early-stage Candida albicans biofilm. *Antimicrob Agents Chemother*. 2002; 46: 3522-31.
  12. Torres SR, Peixoto CB, Caldas DM, Akiti T, Barreiros MGC, Uzeda M, et al. A prospective randomized trial to reduce Candida spp. colonization in patients with hyposalivation. *Braz Oral Res*. 2007; 21: 182-7.
  13. Ellepola ANB, Samaranayake LP. Adjunctive use of chlorhexidine in oral candidosis: a review. *Oral Dis*. 2001; 7: 11-7.
  14. Bobichon H, Bouchet P. Action of chlorhexidine on budding Candida albicans: screening and transmission electron microscopic study. *Mycopathologia*. 1987; 100: 27-35.
  15. D.C. Jagger, A. Harrison. Denture cleansing - the best approach. *Braz Dent J*. 1995;178:413-417
  16. Ntrouka VI, Slot DE, Louropoulou A, Van derWeijden F: The effect of chemotherapeutic agents on contaminated titanium surfaces: a systematic review. *Clin Oral Implants Res* 2011, 22:681-690.
  17. Ntrouka V, Hoogenkamp M, Zaura E, van derWeijden F: The effect of chemotherapeutic agents on titanium-adherent biofilms. *Clin Oral Implants Res* 2011, 22:1227-1234.
  18. Gomes PN, da Silva WJ, Pousa CC, Narvaes EA, Del Bel Cury AA: Bioactivity and cellular structure of Candida albicans and Candida glabrata biofilms grown in the presence of fluconazole. *Arch Oral Biol* 2011, 56:1274-1281.
  19. Nikawa H, Hamada T, Yamashiro H, Kumagai H: A review of in vitro and in vivo methods to evaluate the efficacy of denture cleansers. *Int J Prosthodont* 1999, 12:153-159.
  20. Mouhyi J, Sennerby L, Pireaux JJ, Dourov N, Nammour S, Van Reck J: An XPS and SEM evaluation of six chemical and physical techniques for cleaning of contaminated titanium implants. *Clin Oral Implants Res* 1998, 9:185-194.
  21. Chandra J, Kuhn DM, Mukherjee PK, Hoyer LL, McCormick T, Ghannoum MA. Biofilm formation by the fungal pathogen Candida albicans: Development, architecture and drug resistance. *J Bacteriol*. 2001; 183: 5385-94.
  22. Lamfon H, Porter SR, McCullough M, Pratten J. Susceptibility of Candida albicans biofilms grown in a constant depth film fermentor to chlorhexidine, fluconazole and miconazole: a longitudinal study. *J Antimicrobial Chemother*. 2004; 53: 383-5.
  23. Makihira S, Nikawa H, Tamagami M, Hamada t, Samaranayake LP. Differences in Candida albicans adhesion to intact and denture type I collagen in vitro. *Oral Microbiol Immunol*. 2002; 17: 129-31.
  24. Meiller TF, Kelley JI, Jabra-Rizk MA, DePaola LG, Baqui A, Falker WA. In vitro studies of the efficacy of antimicrobials against fungi. *Oral Surg Oral Med Oral Pathol Oral RadiolEndod*. 2001; 91: 663-70.
  25. Fernanda Campos Machado, Maristela Barbosa Portela, Amanda Carneiro da Cunha, Ivete Pomarico Ribeiro de Souza, Rosângela Maria de Araújo Soares, Gloria Fernanda Barbosa de Araújo Castro. Antifungal activity of chlorhexidine on Candida spp. Biofilm. *Rev Odontol UNESP, Araraquara*, 2010; 39(5): 271-275
  26. MacNeill S, Rindler E, Walker A, Brown AR, Cobb CM. Effects of tetracycline hydrochloride and chlorhexidinegluconate on Candida albicans. An in vitro study. *J ClinPeriodontol*. 1997; 24: 753-60.
  27. Franco Neto CA, Parolo CCF, Rösing CK, Maltz M. Comparative analysis of the effect of two chlorhexidinemouthrinses on plaque accumulation and gingival bleeding. *Braz Oral Res*. 2008; 22: 139-44.

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